

Bioengineered approaches for repairing nerve injury

John Haycock

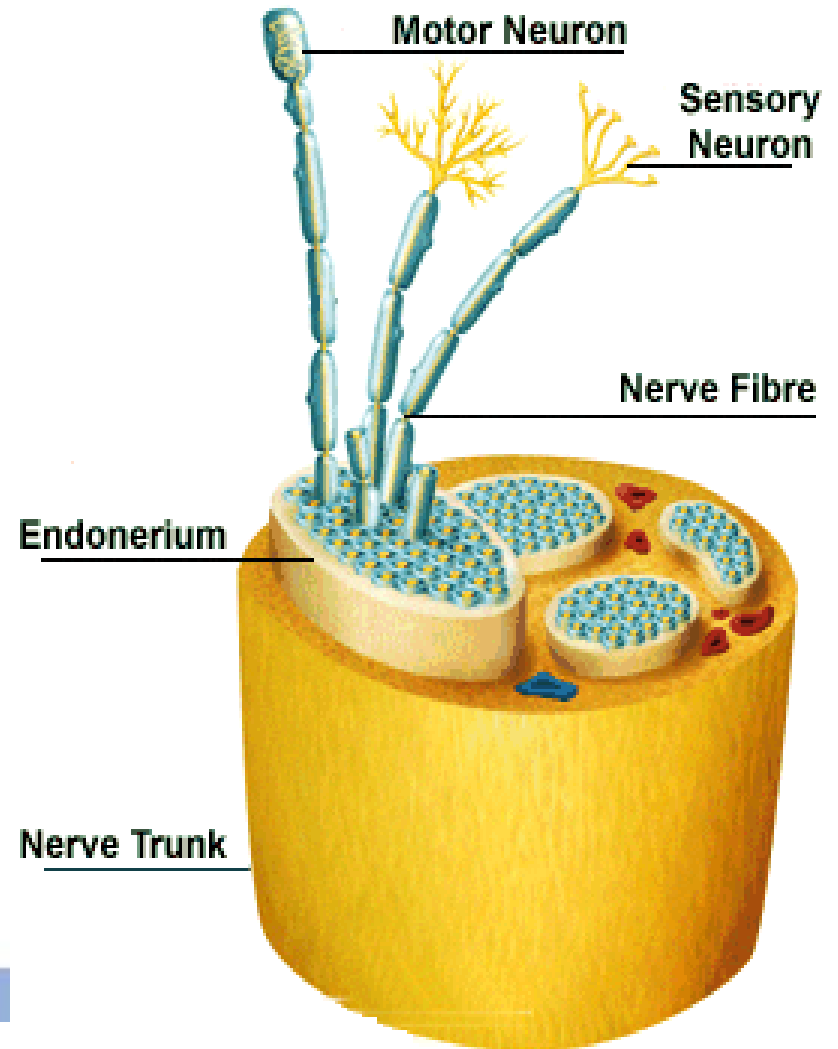
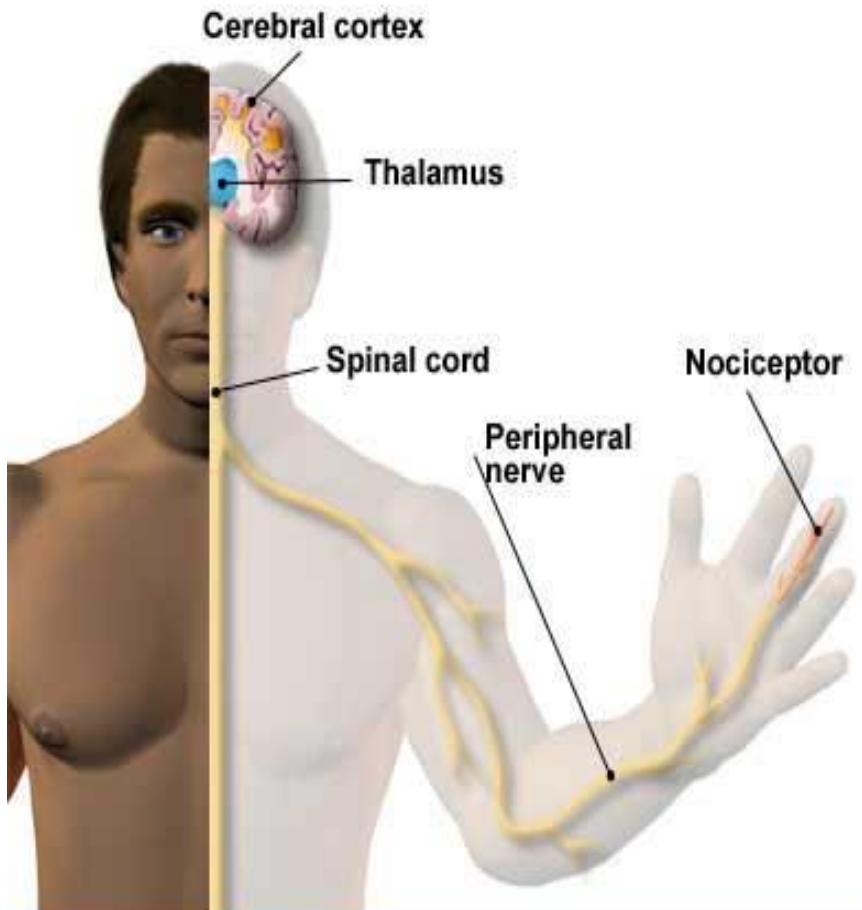
Professor of Bioengineering

Head of Materials Science and Engineering
University of Sheffield, UK.



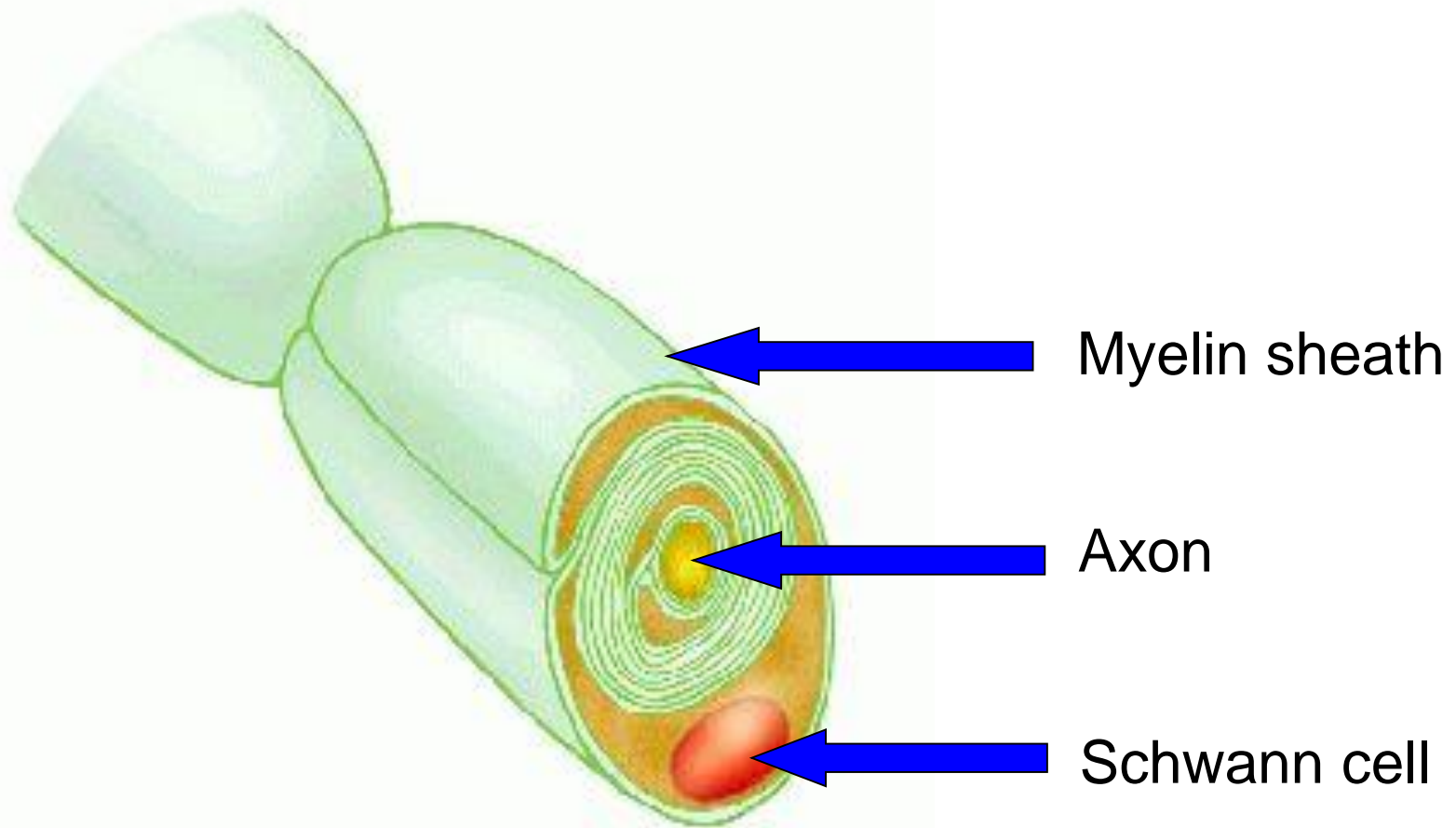


Peripheral nerve





Peripheral nerve axons

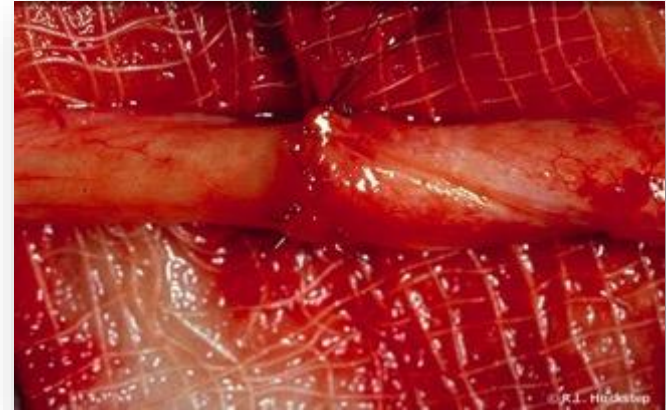




Three main clinical strategies to repair gap injuries

1. Suturing together proximal and distal ends

- + 'Clean' transection injury
- Tension in sutures



2. Autografting

- + Good reinnervation
- Donor site morbidity



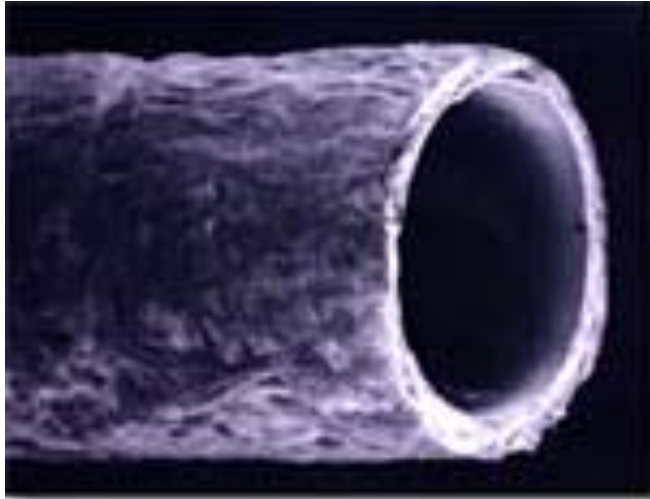
3. Nerve guidance conduits

- + Biocompatible materials
- Primitive design
- Limited regeneration

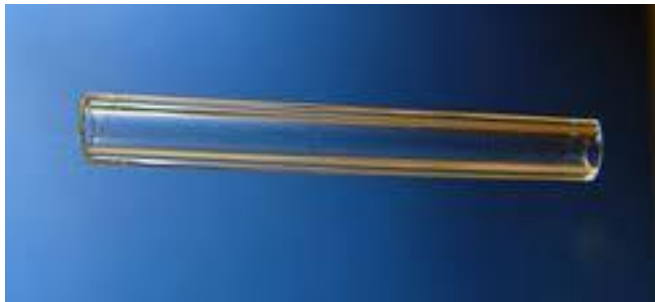




Nerve Guidance Channels



- Collagen - Integra Life Sciences NeuraGen™ nerve guide
- Silicone - SaluMedica's SaluBridge™ nerve cuff

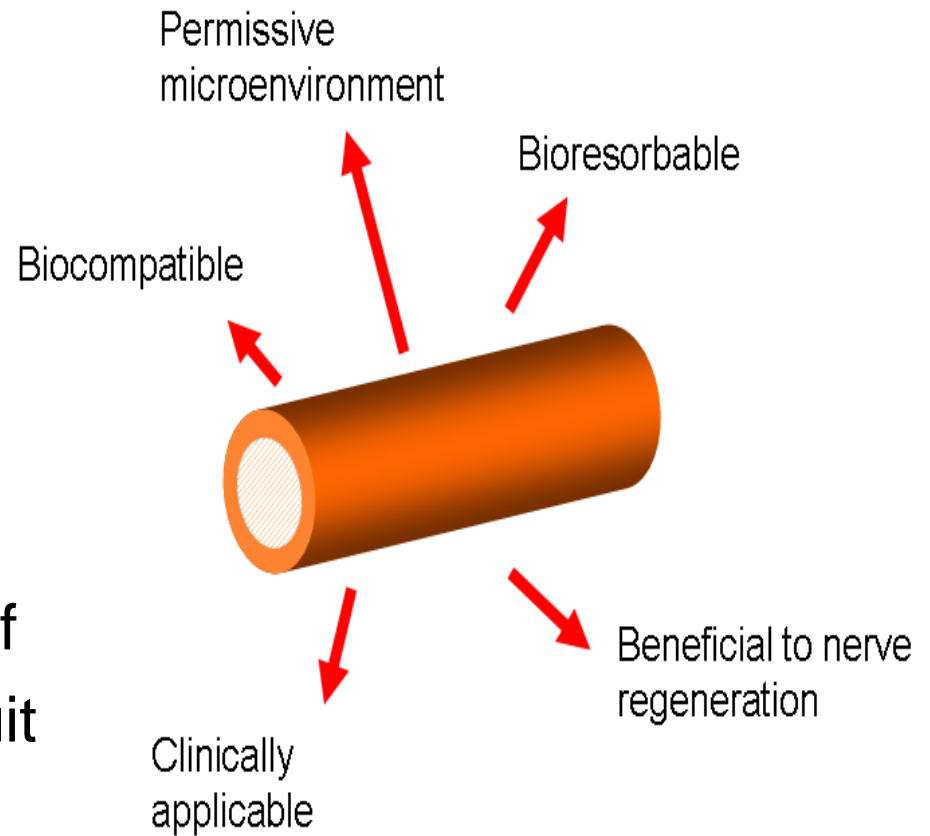


- PGA – Neurotube (Synovis)
- PLLA/PCL – Neurolac (Polyganics)

Bell JHA and Haycock JW (2012). Next generation nerve guides - materials, fabrication, growth factors and cell delivery. *Tissue Engineering* 18(2):116-28

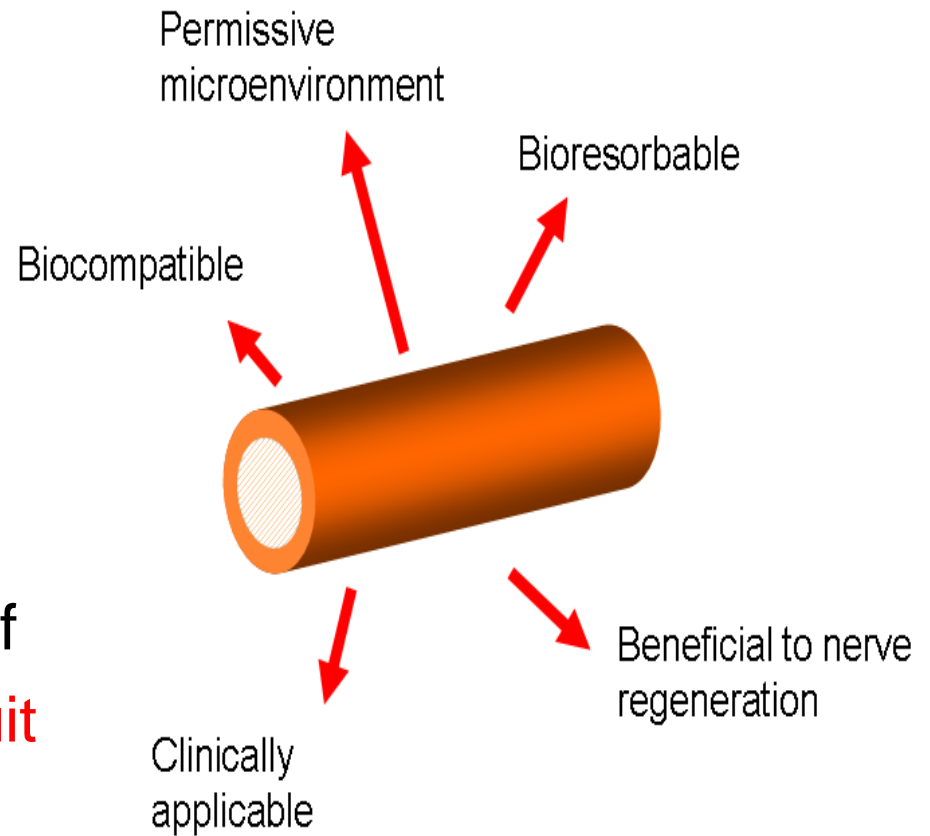
Present strategies for repairing peripheral nerve

- To increase regeneration distance
- To improve extent and effectiveness of reinnervation
- Involves a combination of
 - 1) Nerve guidance conduit
 - 2) Schwann cells



Present strategies for repairing peripheral nerve

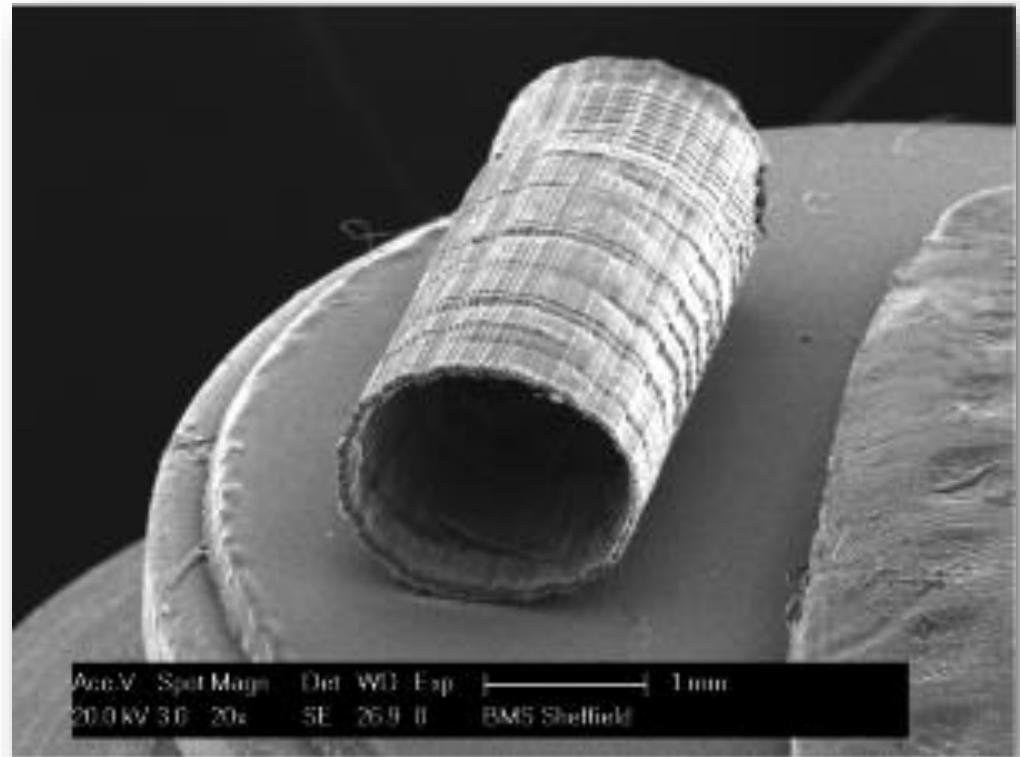
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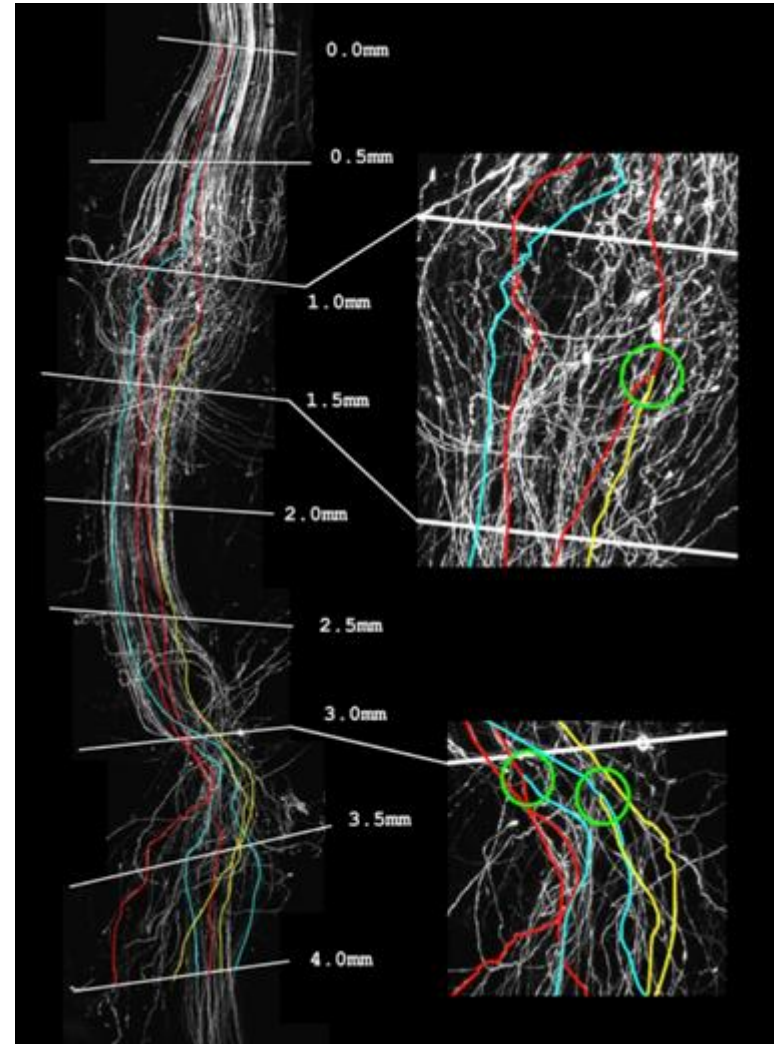
Making a scaffold precisely

Micro-stereolithography

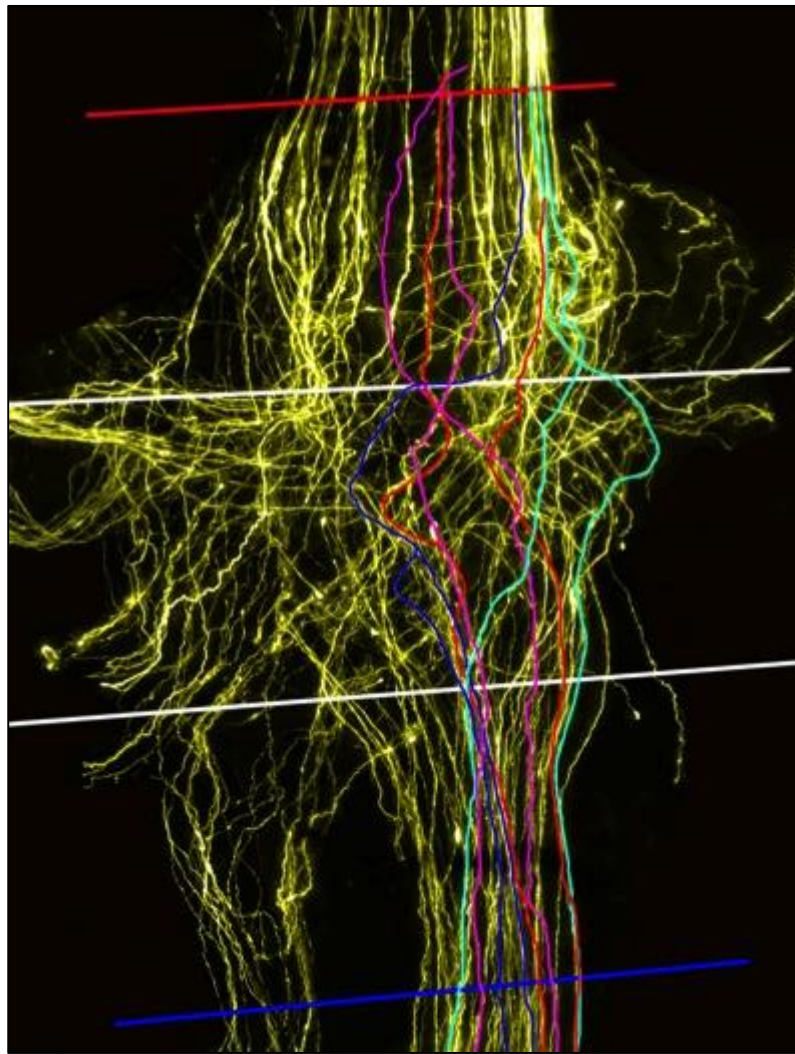
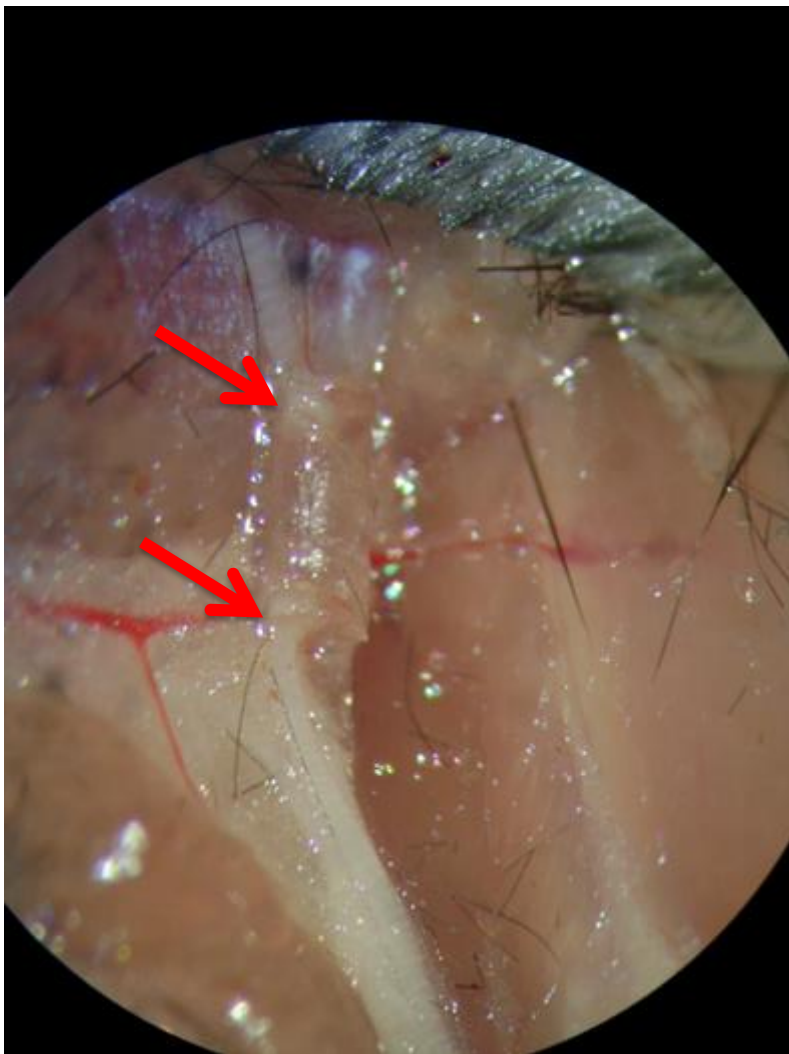
1. Manufacture of NGCs - PEG, PLA, PCL, PGS
2. Incorporate internal structure within the tube to improve regeneration



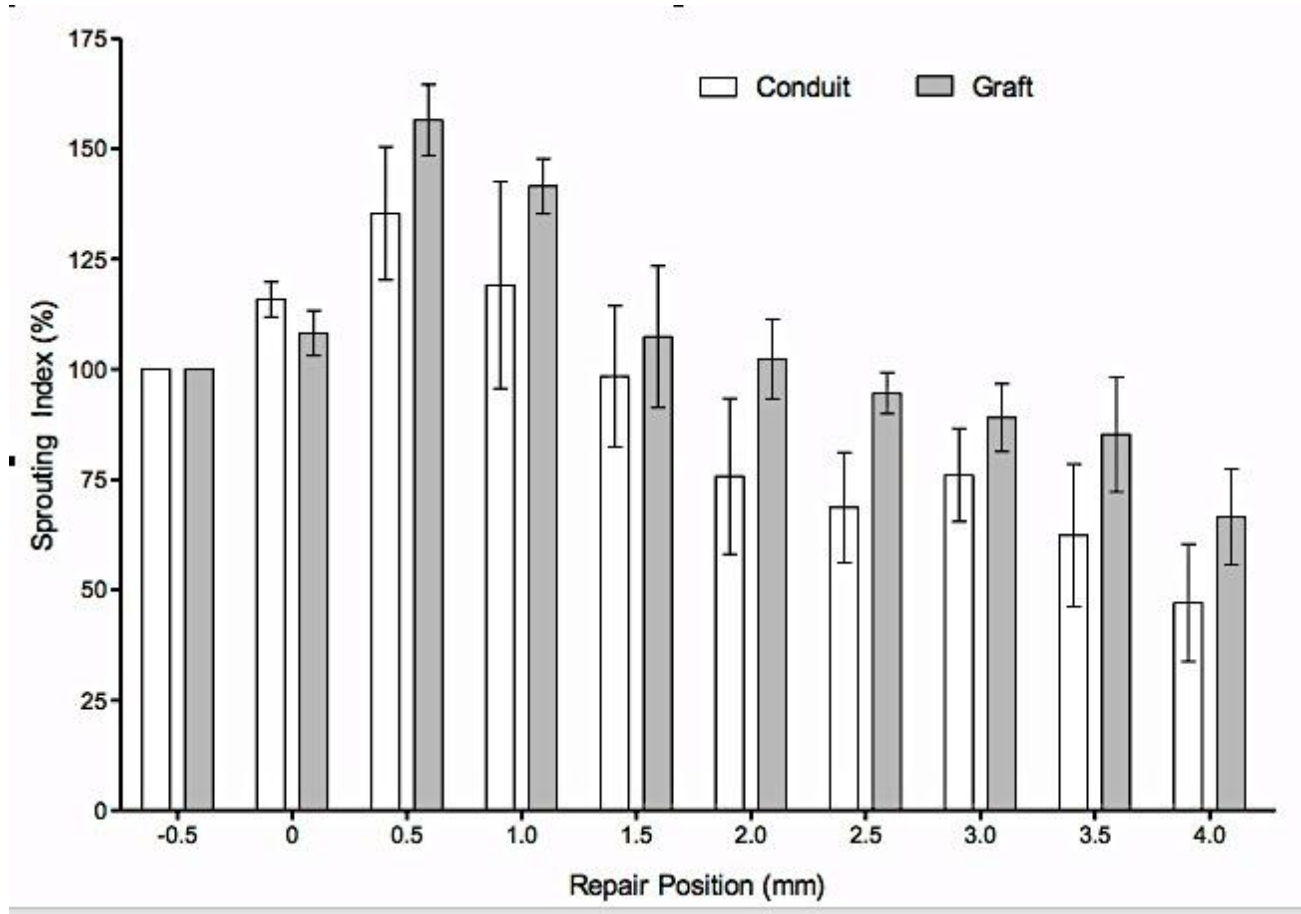
YFP mouse – 3mm common fibular nerve injury model



YFP mouse – 3mm common fibular nerve injury model



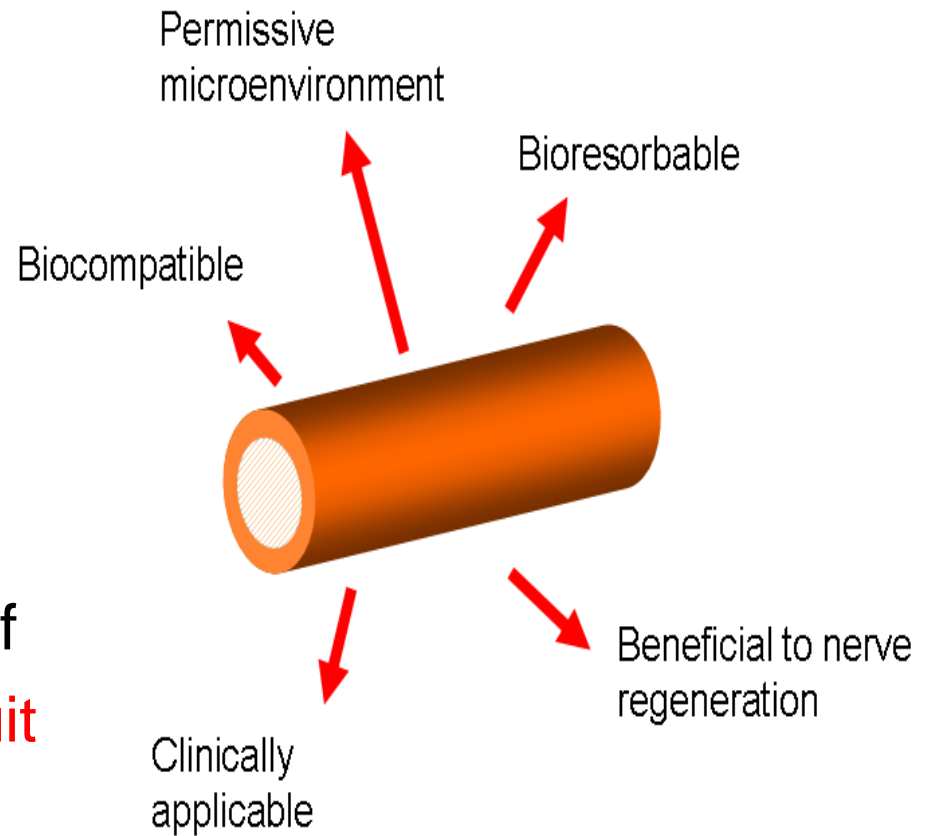
Graft versus nerve conduit repair



Pateman C, Harding A, Glen A, Taylor C, Christmas C, Robinson P, Rimmer S, Boissonade F, Claeysens F, **Haycock JW**. (2015) Nerve guides manufactured from photocurable polymers to aid peripheral nerve repair. *Biomaterials* 49, 77–89.

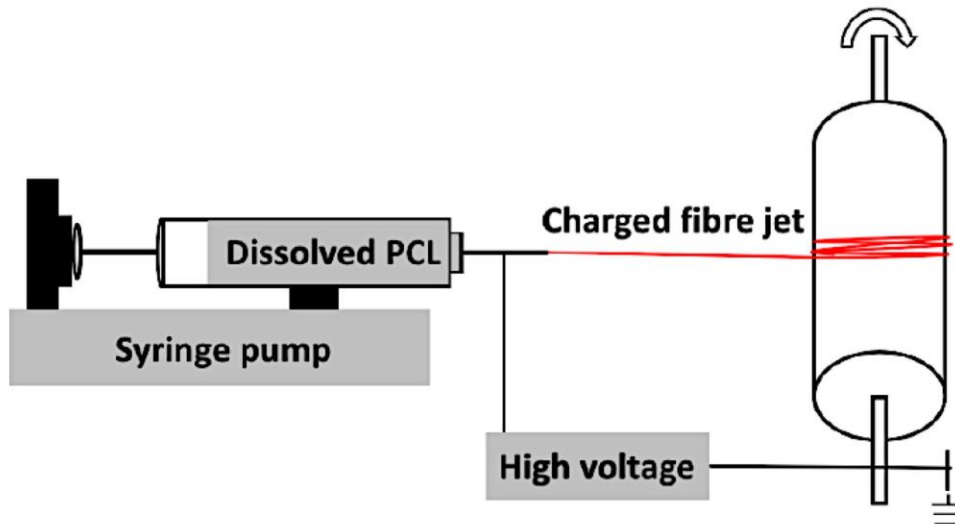
Present strategies for bioengineering peripheral nerve

- To increase regeneration distance
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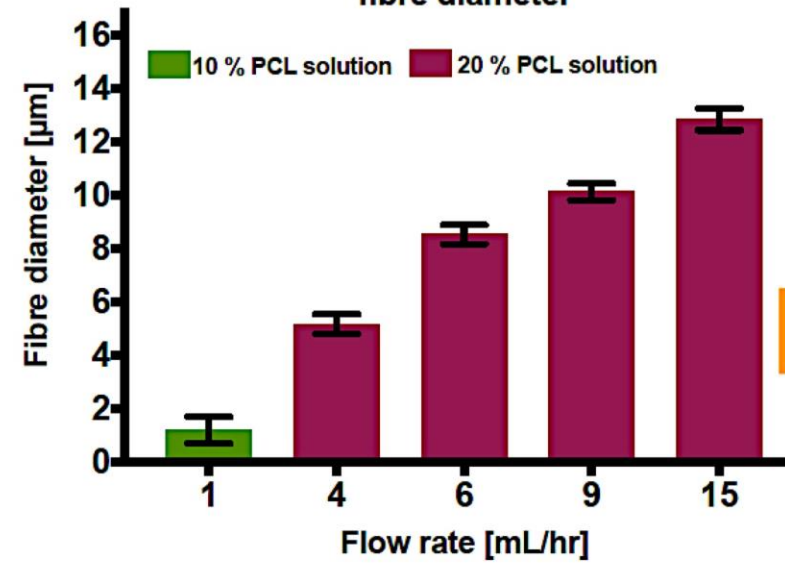


3D In vitro assessment model

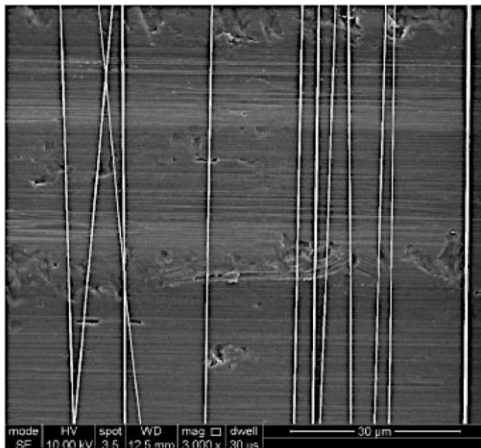
Electrospinning of aligned PCL microfibres



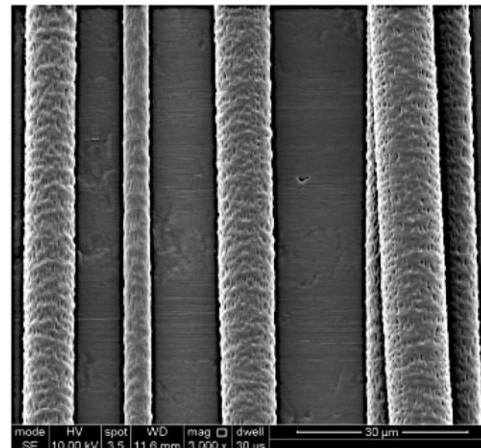
Increasing flow rates increase PCL fibre diameter



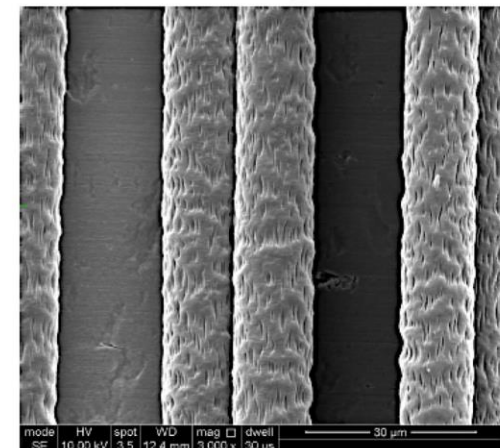
1 µm



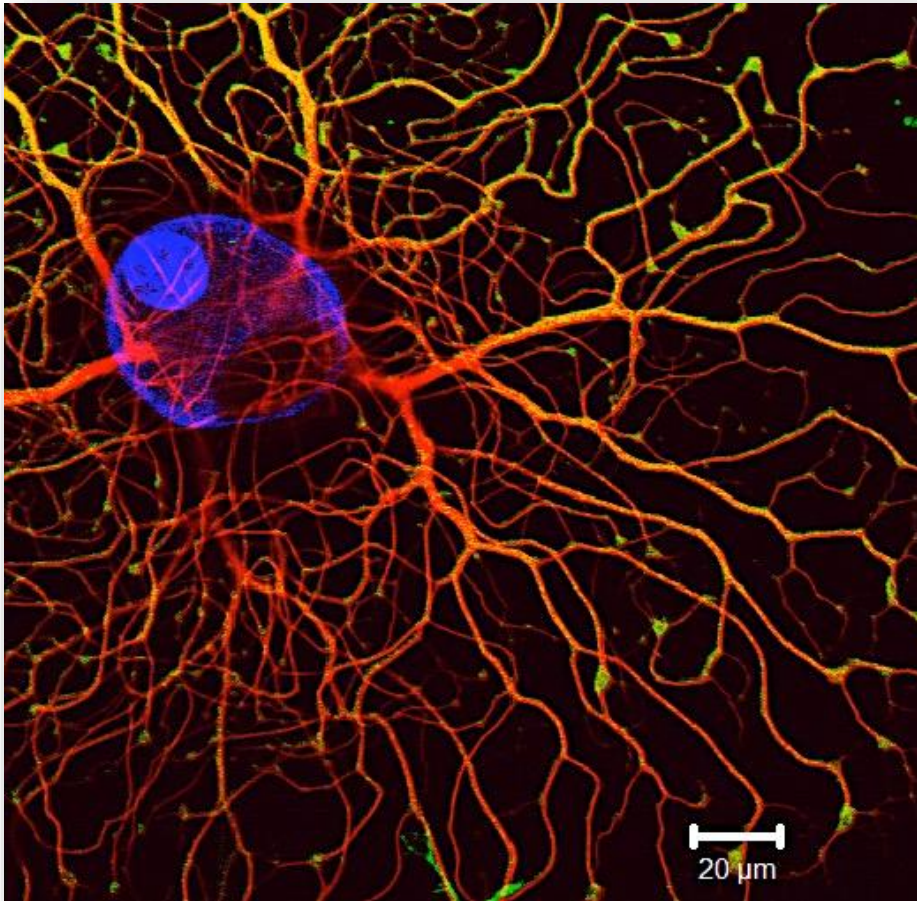
8 µm



13 µm



Dorsal Root Ganglion cultures

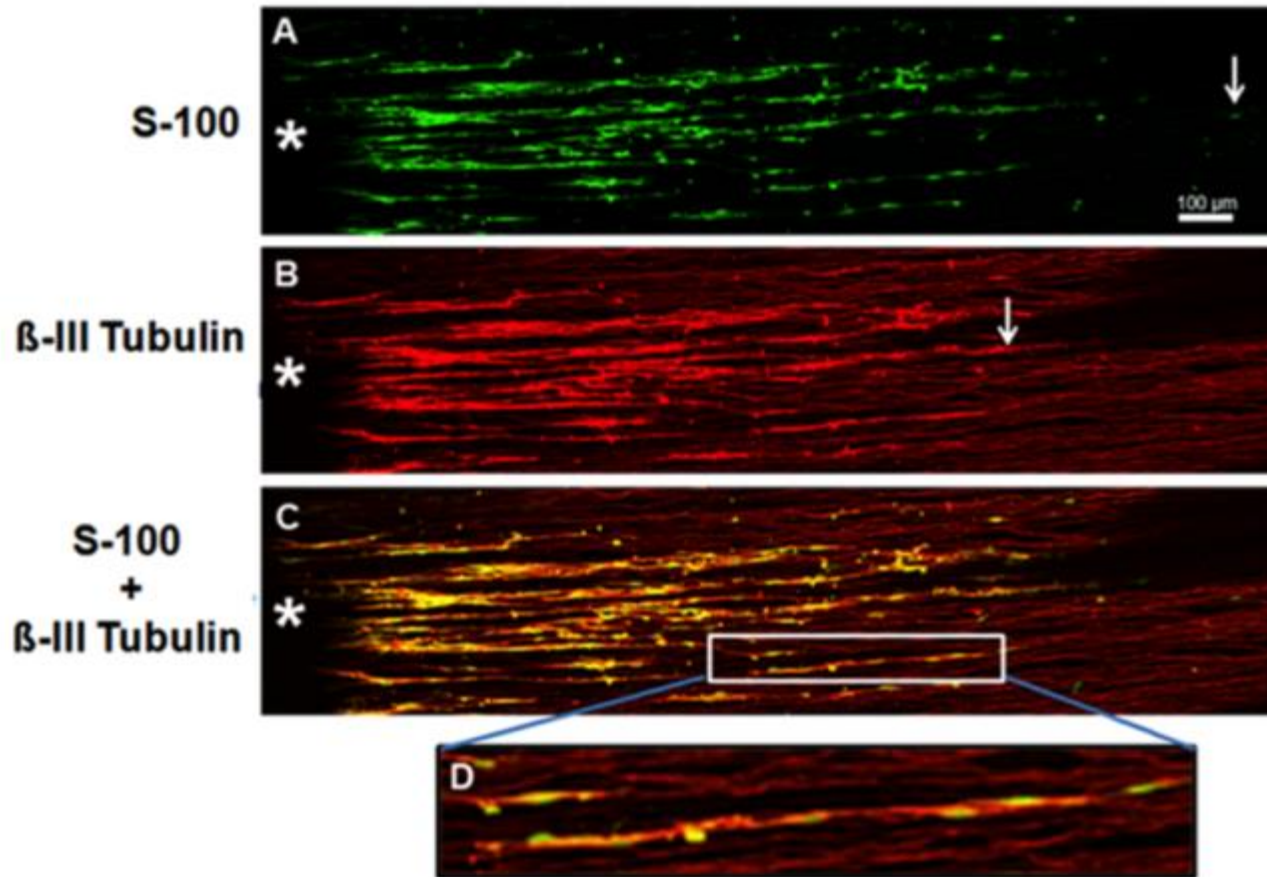


- On flat culture surfaces DRG neurites form a highly connected but disorganised network
- Nuclei
- β -tubulin-III
- S100 β
- Can DRG neurites and Schwann cells be organised to resemble a peripheral nerve?

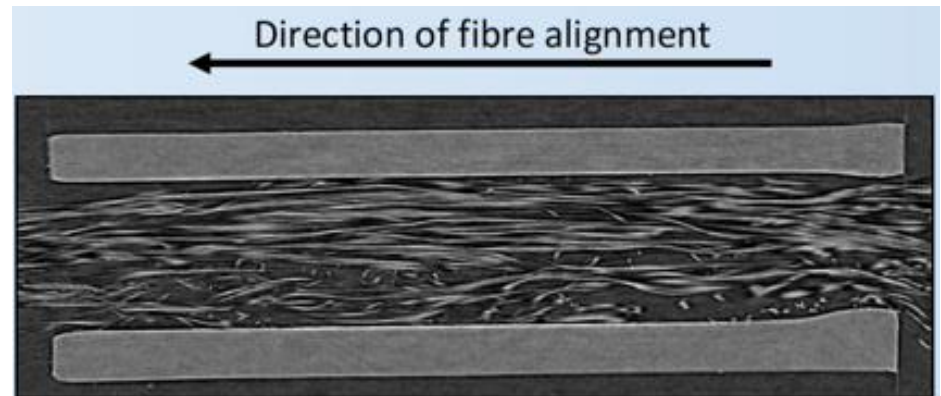
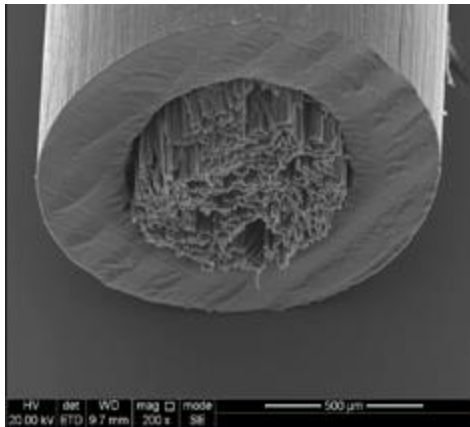
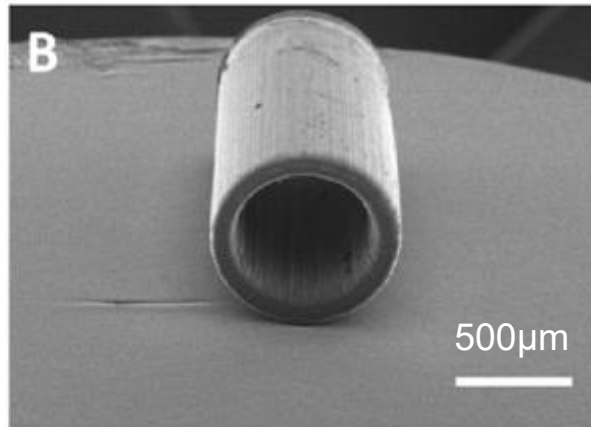


PCL aligned fibre scaffolds for organised growth of DRG neurites and Schwann cells

1 μm PCL aligned fibres

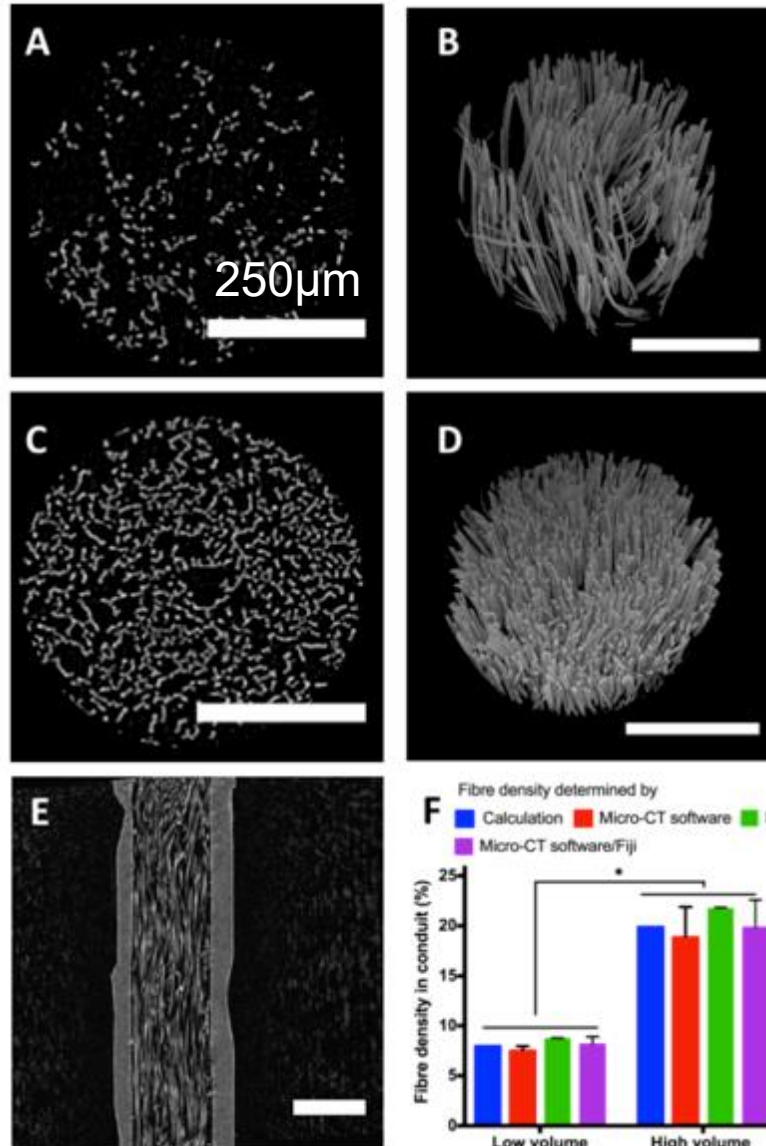


Photocurable poly(caprolactone) conduit + poly(caprolactone) fibres





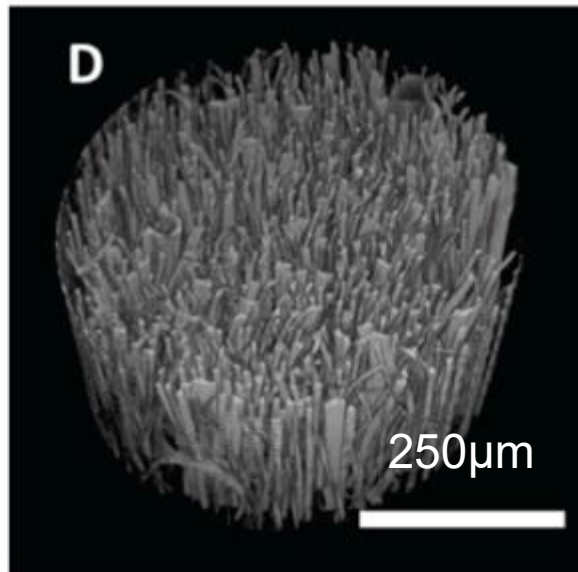
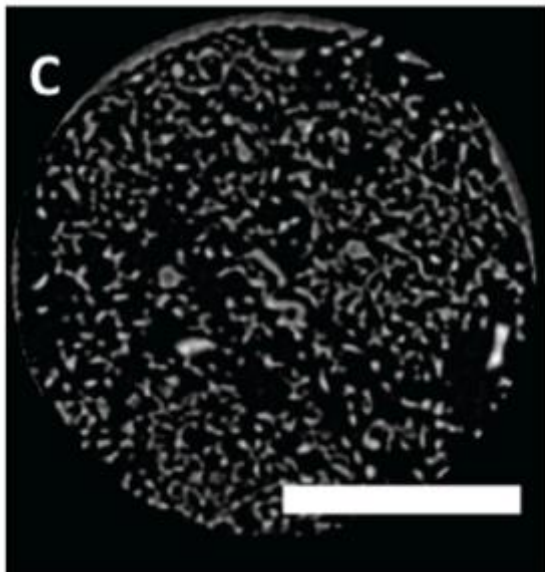
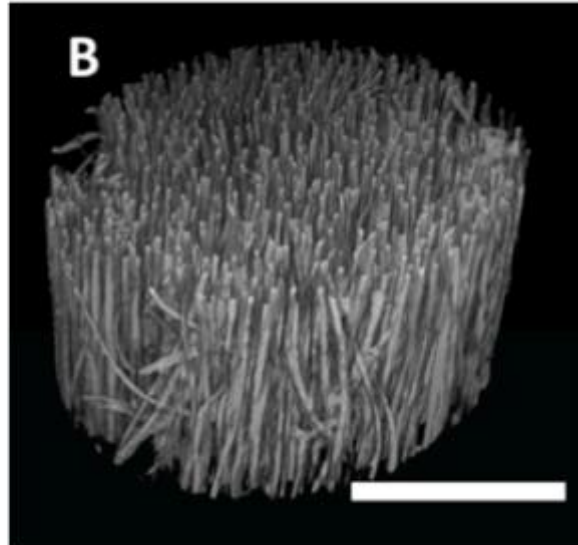
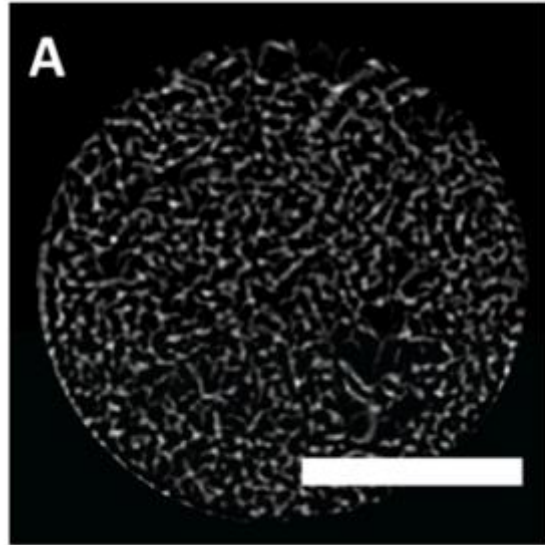
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Micro CT imaging of NGCs

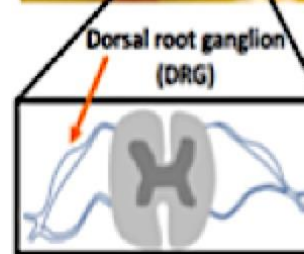
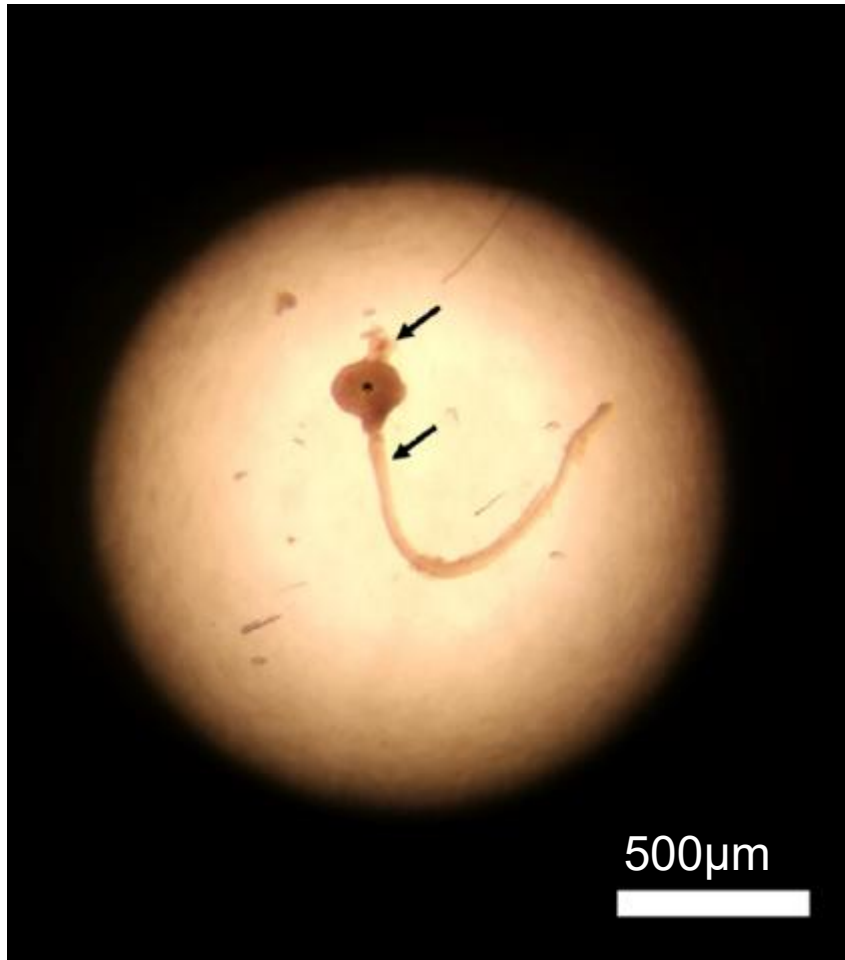


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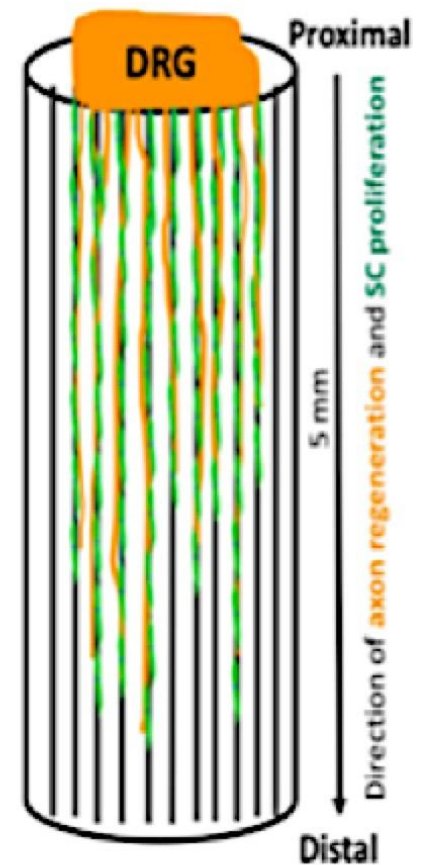


Micro CT
imaging of
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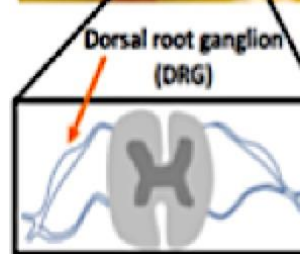
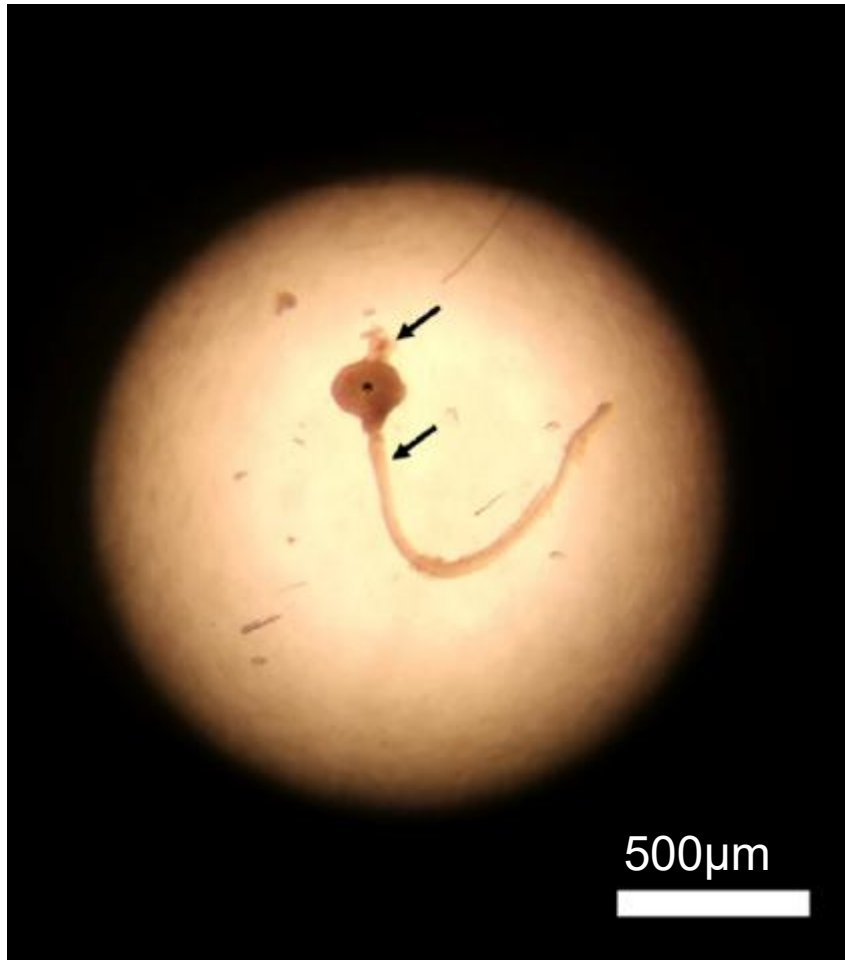
3D In vitro assessment model E12 chick dorsal root ganglion



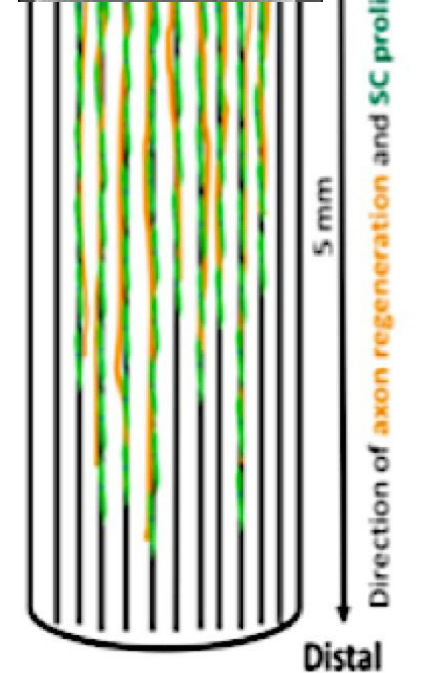
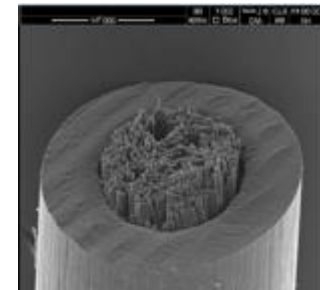
Culture insert for NGC test devices



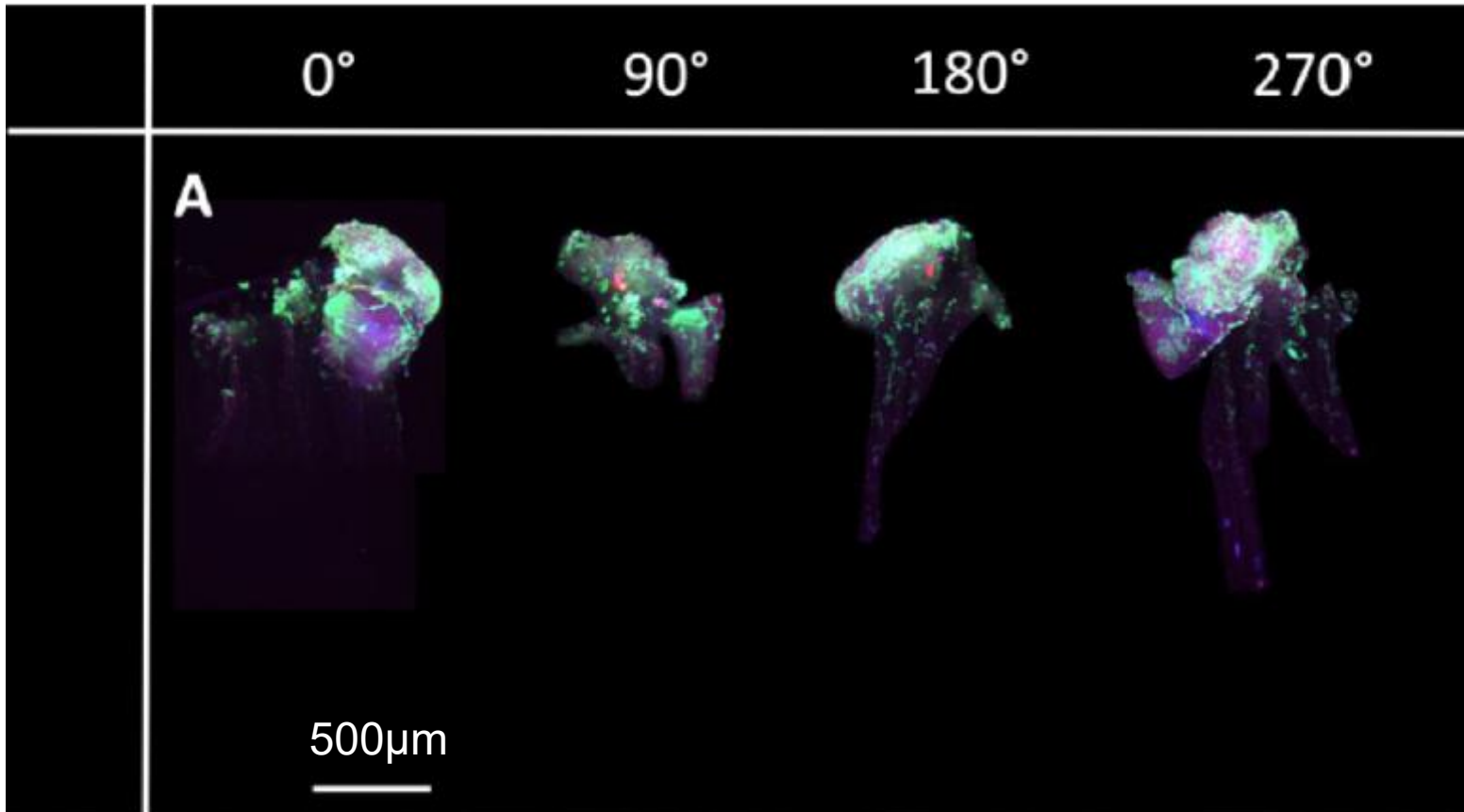
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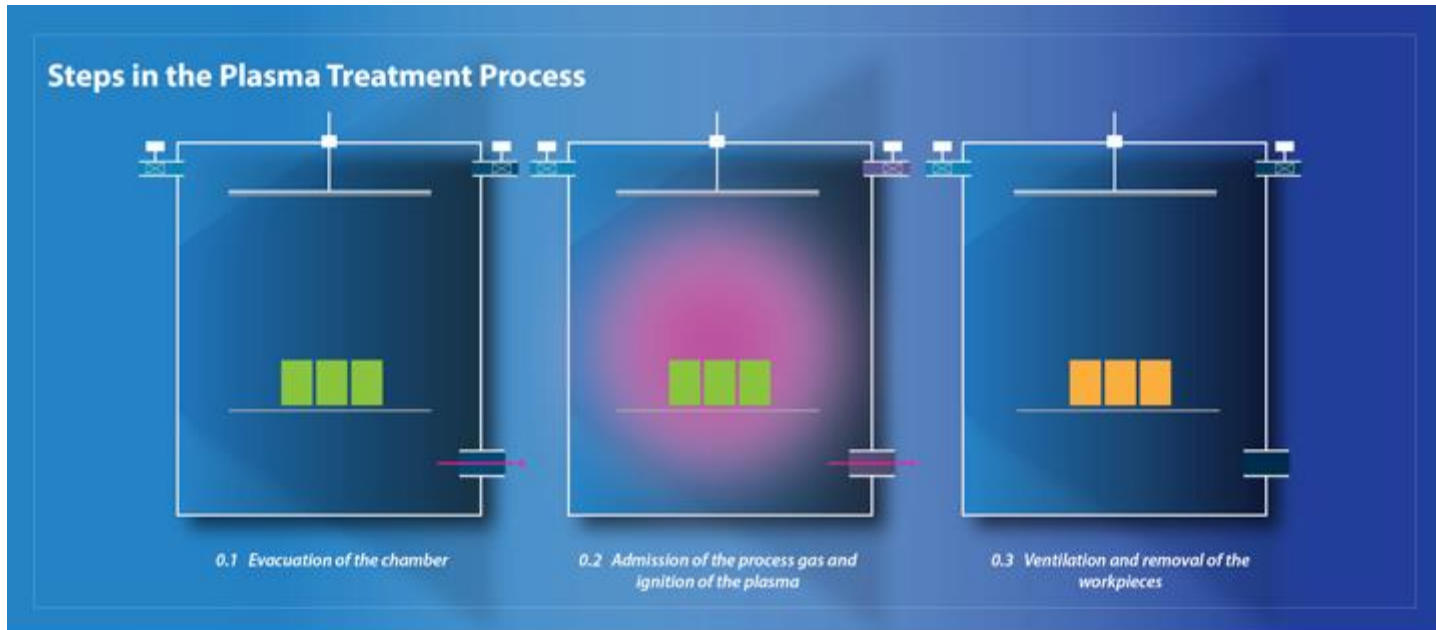


3D In vitro assessment model E12 chick dorsal root ganglion



Light sheet microscopy (Zeiss Z1)
Blue = nuclei / Green = β III tubulin / Red = S100 β

Surface modification of PCL fibres – air plasma



Question – Does air plasma surface deposition of PCL fibres change surface energy, elemental composition and adhesion / growth of neurons in a nerve guide device?

Diener Electronic commercial plasma system (model ZEPTO, chamber volume: 2.6L) with connected pump (Pfeiffer Vacuum Technology AG), 50 W (40 kHz) 0.4 mbar for 60 s.

Air plasma surface treatment of PCL fibres

X-ray photoelectron spectroscopy (XPS)

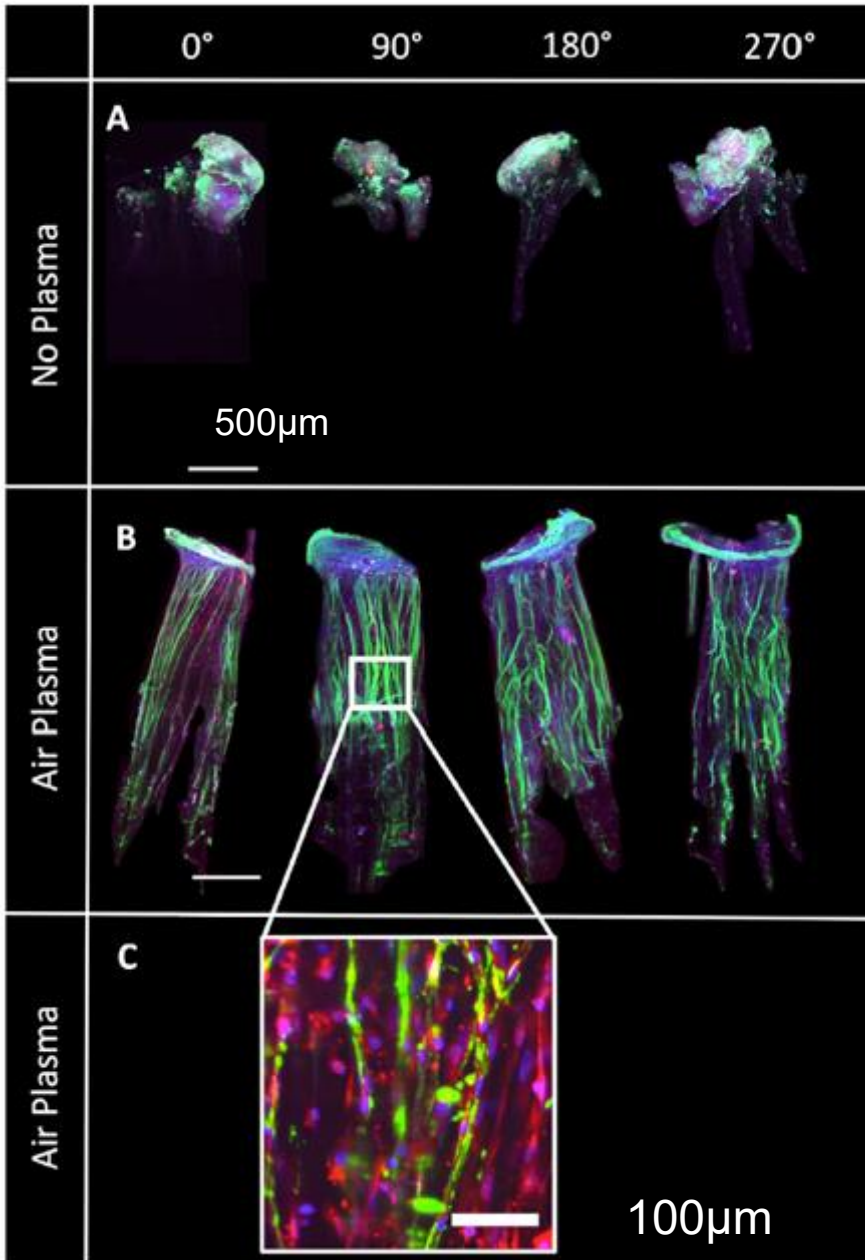
Sample	Wide scan (%)			Narrow scan						
				C1s region (%)					O1s region (%)	
	C	O	N	C-C	C-C-O	C(O)-O-C	COO	C=O	C=O	C(O)-O
FB - #1	79.6	20.5	0	45.1	12.3	12.6	9.7	0	10.5	9.9
FB - #2	82.8	17.2	0	50.3	13.3	11.6	7.6	0	9.4	7.8
FB - #3	79.3	20.7	0	40.6	14.8	13.6	10.2	0	10.2	10.6
FB + #1	78.1	17.6	0.9	29.5	25.1	13.1	5.6	4.9	10.6	7.0
FB + #2	78.4	17.8	1.2	29.3	26.1	13.2	5.5	4.4	10.8	7.0
FB + #3	79.1	16.8	1.2	29.8	26.7	13.3	5.4	3.9	10.2	6.6

+ = Plasma

- = No Plasma

= Position on the sample (1 = top, 2 = middle, 3 = bottom)

FB = fibre bundle



Air plasma surface treatment of PCL fibres

E12 Chick DRG

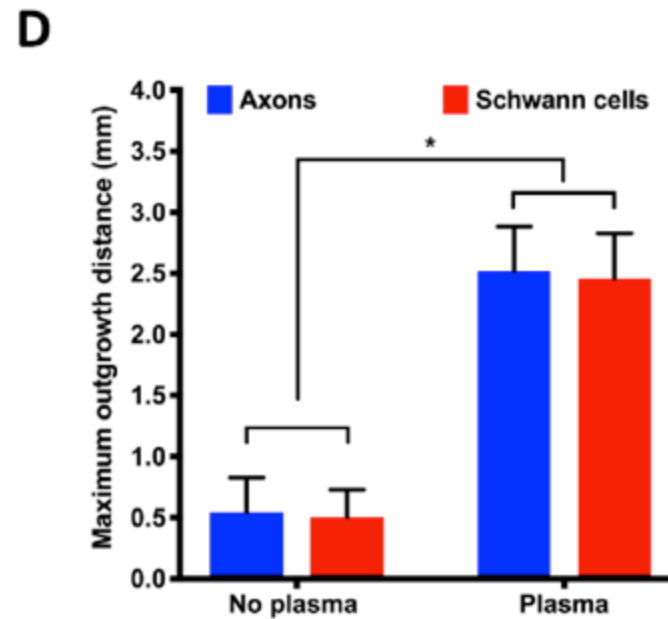
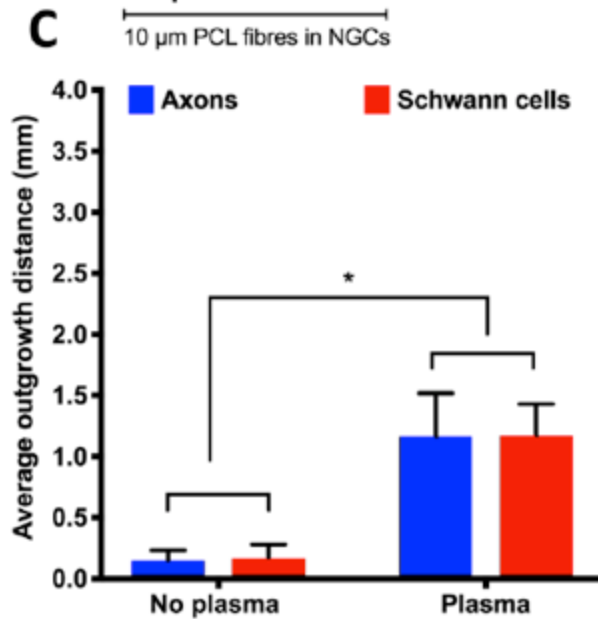
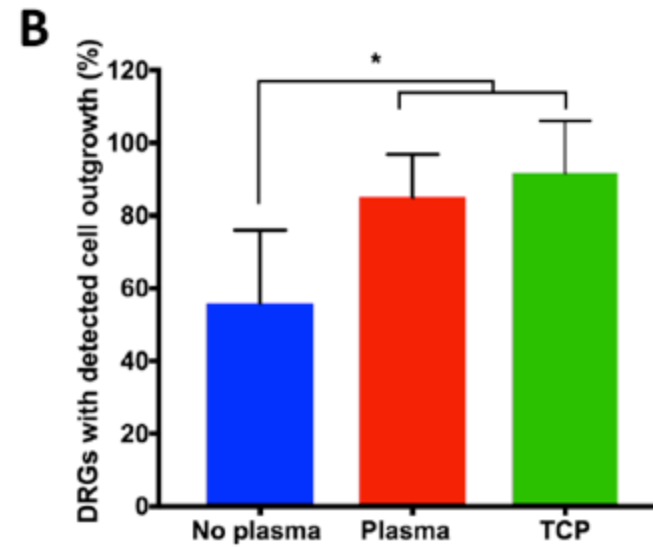
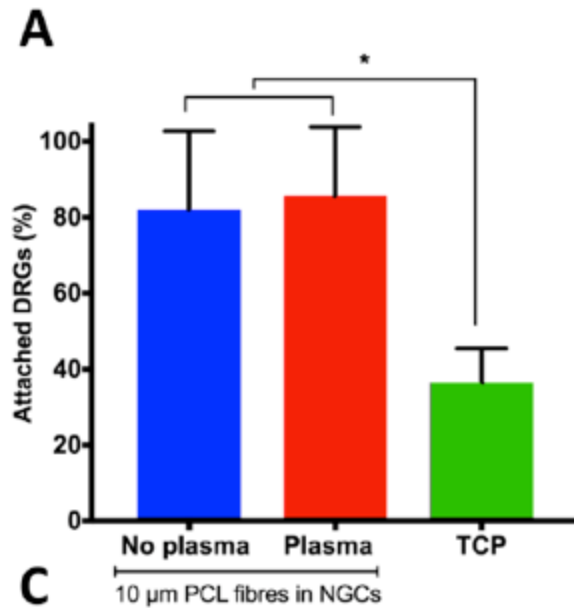
Light sheet microscopy (Zeiss Z1)

Blue = nuclei
Green = β III tubulin
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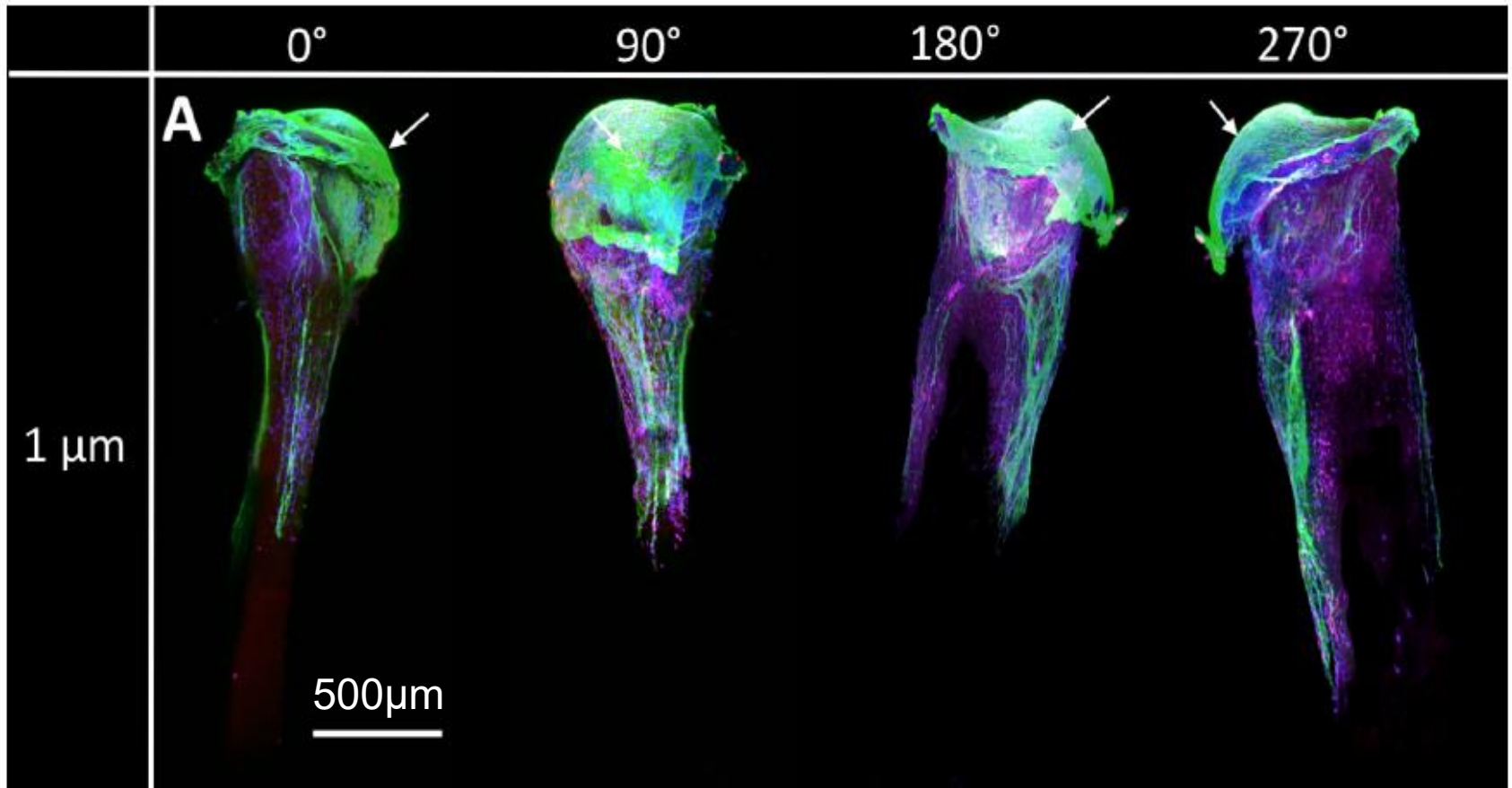
3D In vitro assessment model

E12 Chick DRG





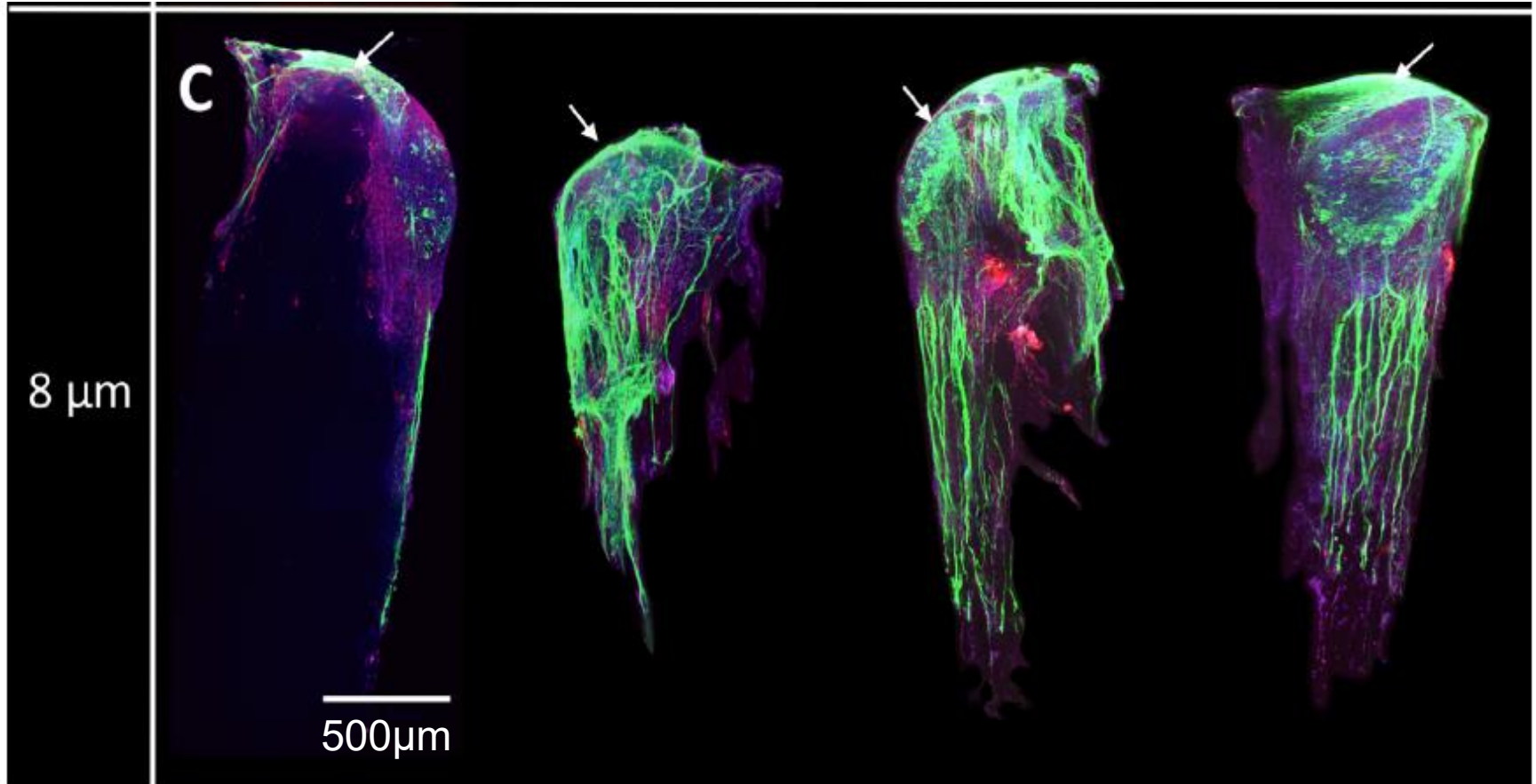
3D In vitro assessment model E12 Chick DRG – PCL fibre diameter



Light sheet microscopy (Zeiss Z1)
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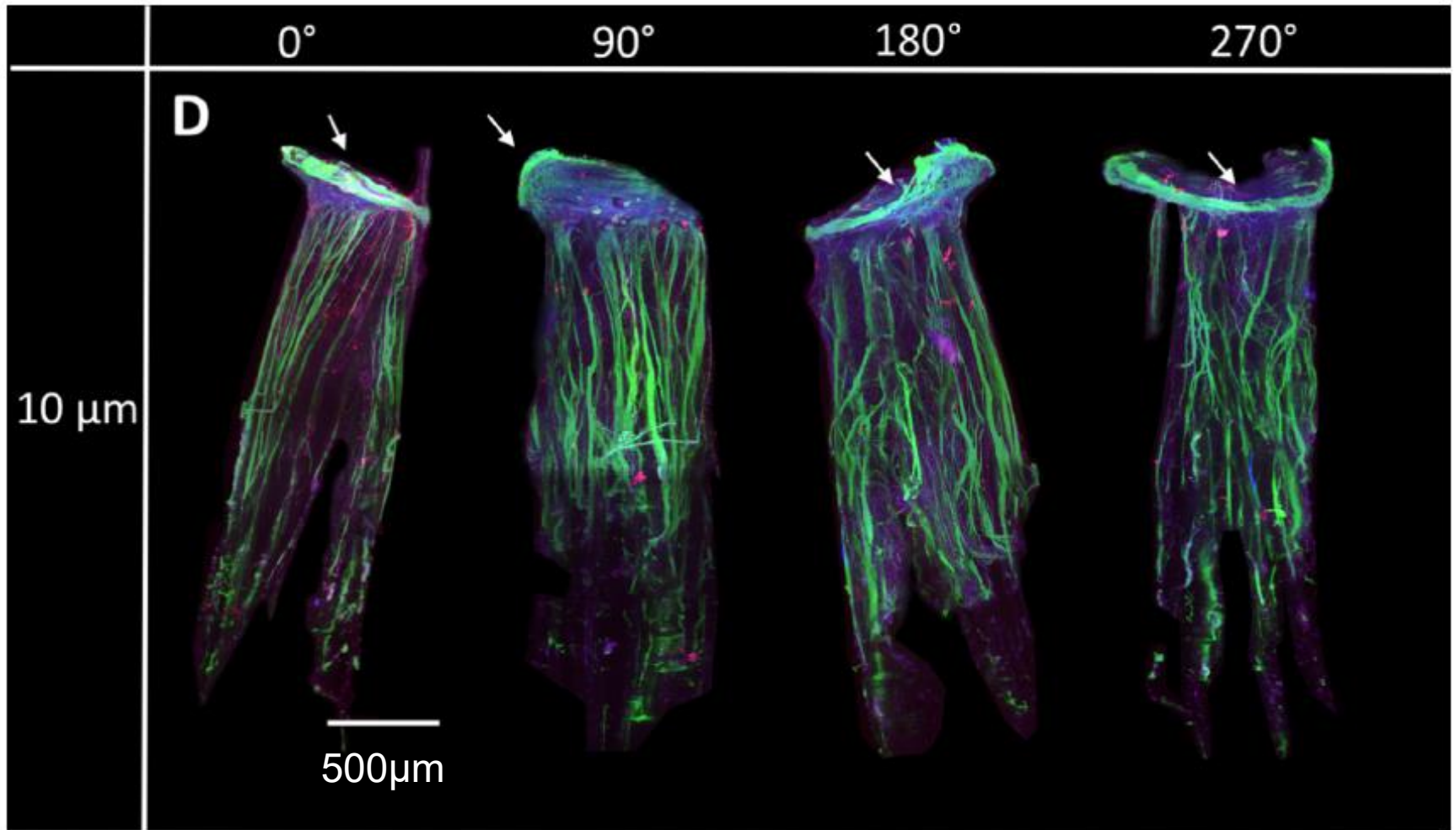
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3D In vitro assessment model

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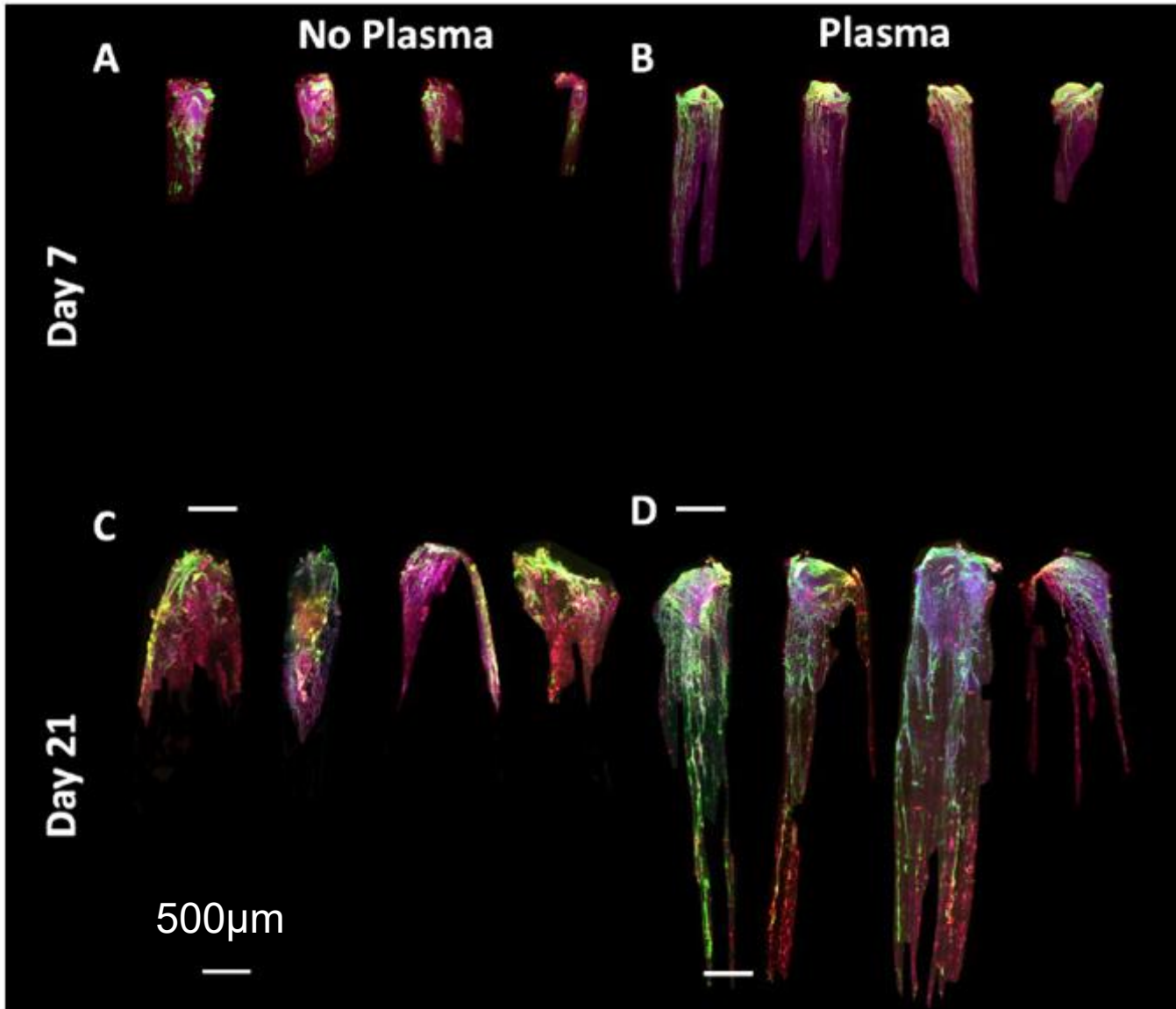


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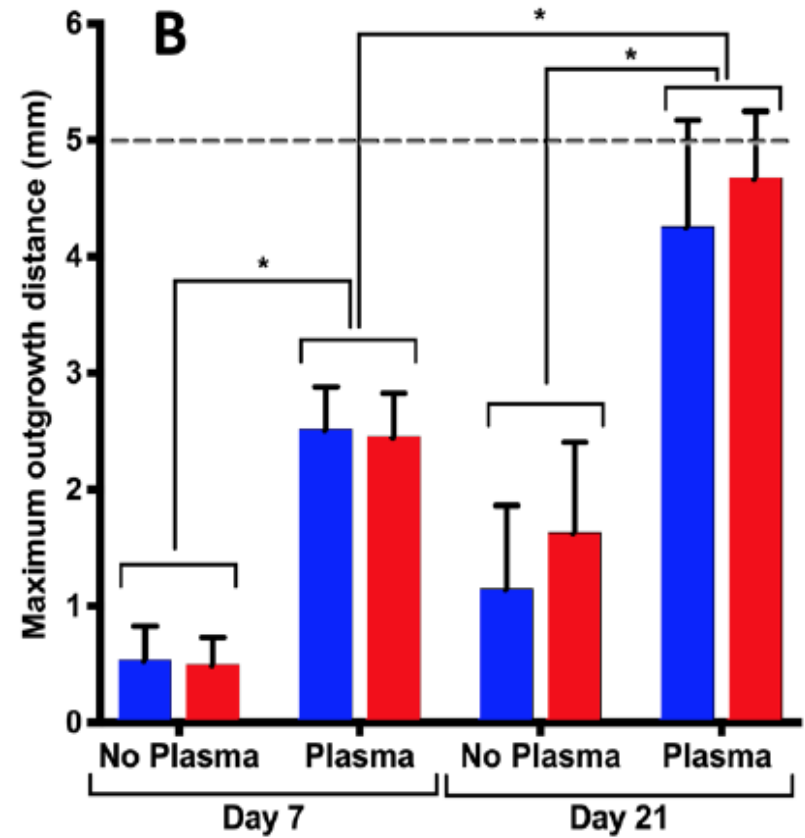
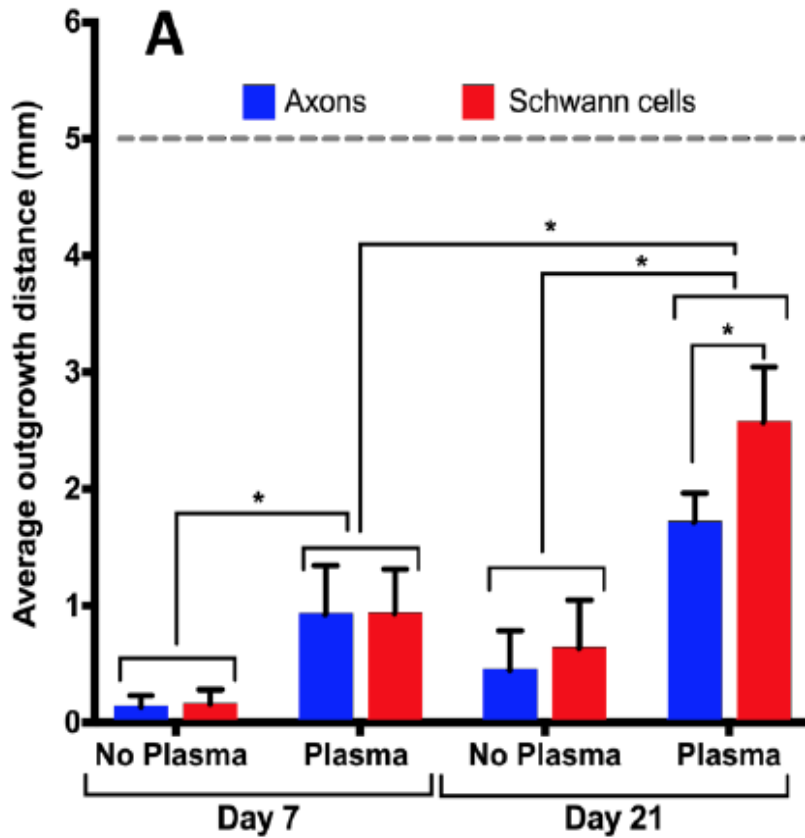
E12 Chick DRG – 7d and 21d (10µm fibres)

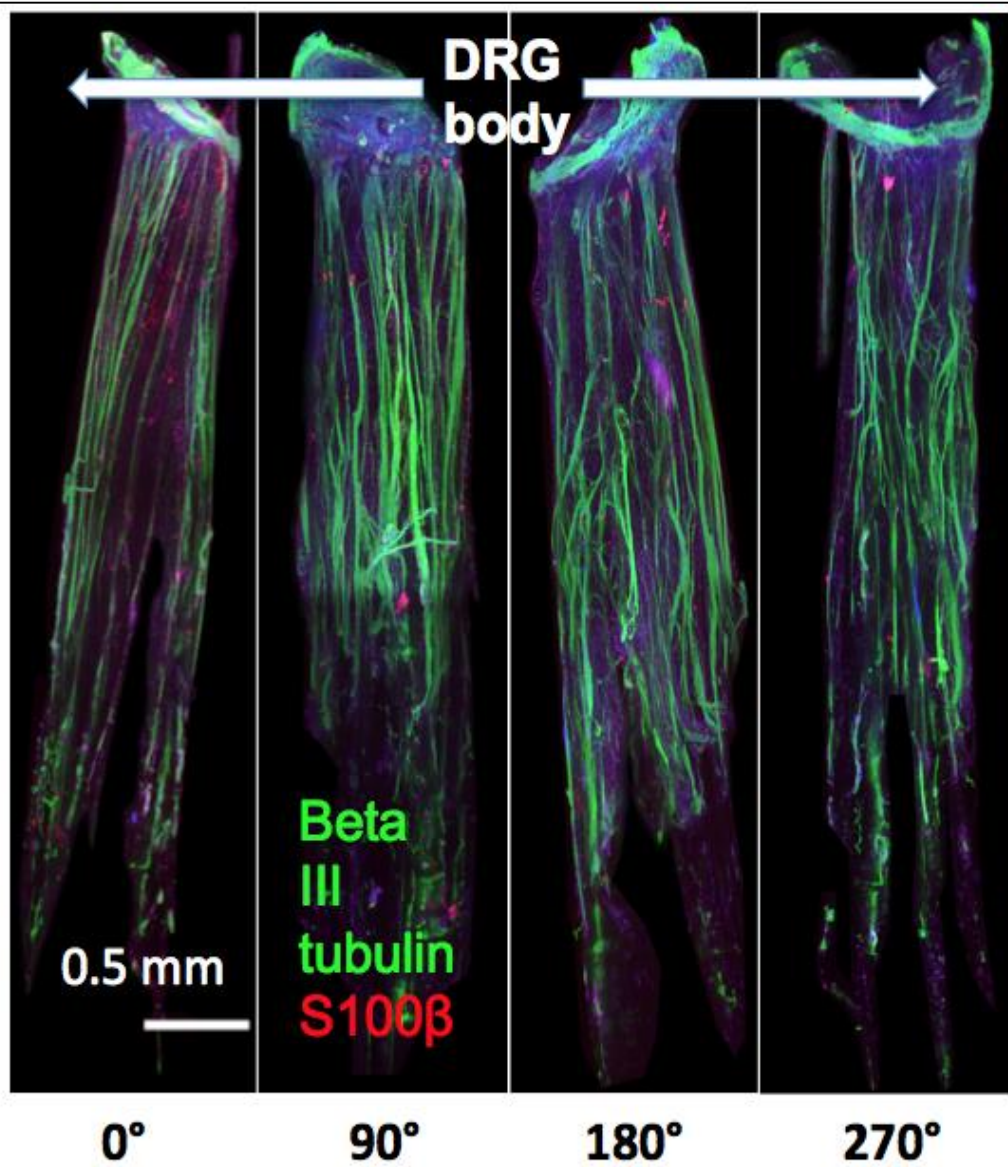
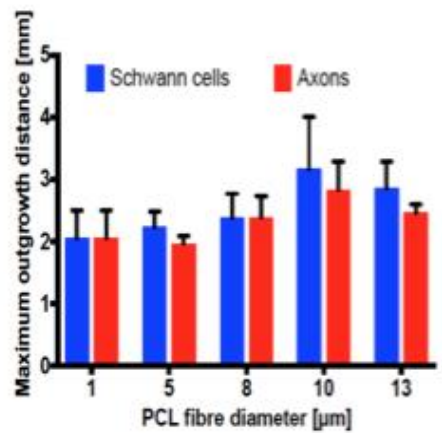
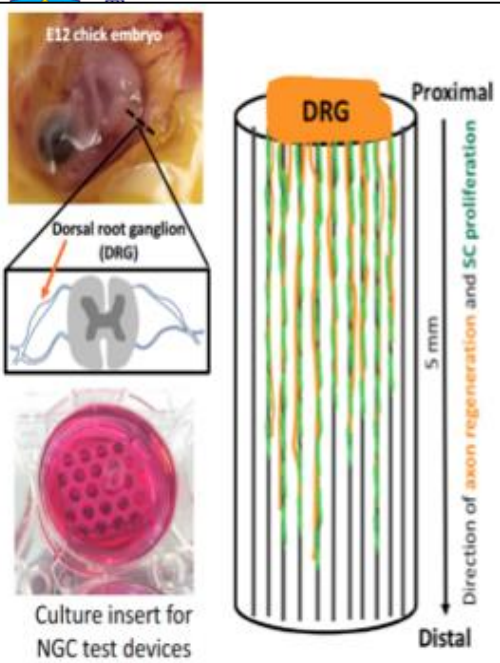




3D In vitro assessment model

E12 Chick DRG – 7d and 21d



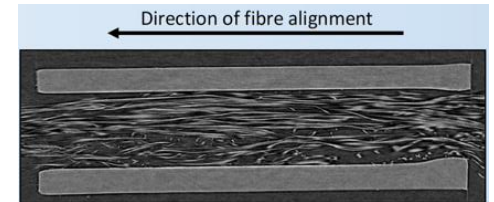
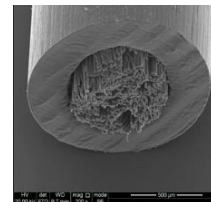
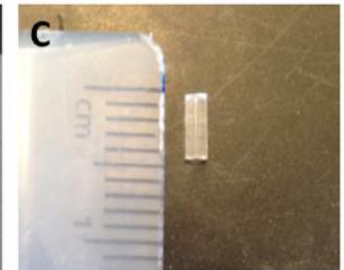
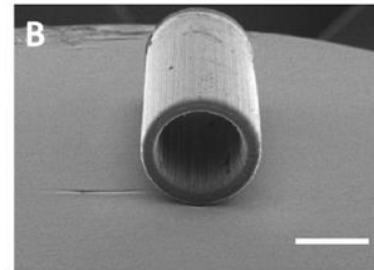
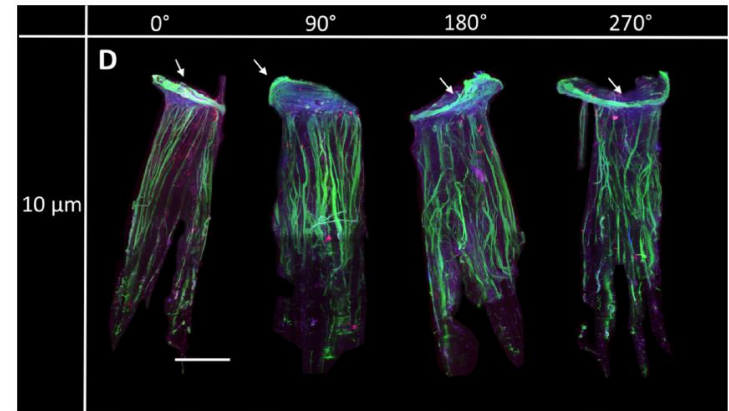


Direction of axon growth and Schwann cell migration over 7 days

Behbehani M, Glen A, Taylor CS, Schumaker A, Claeysens F, Haycock JW (2018) Pre-clinical evaluation of advanced nerve guide conduits using a novel 3D in vitro testing model. *Int J Bioprinting* 4(1) 1-12.

Conclusions

- 3D printed nerve guides made from PEG, and biodegradable PCL, PGS
- Support nerve repair *in vivo*
- Aligned internal fibre devices optimised
- Validated by microCT
- DRG models + light sheet
- Surface modification improves guided nerve growth
- Natural material NGC programme funded by Pak-UK fellowship IRC-BM / COMSATs – Dr Ather Farook Khan
- Study completed and ready to file patent for translation + commercialization



Recent + key publications

- Behbehani M, Glen A, Taylor CS, Schumaker A, Claeysens F, Haycock JW (2018) Pre-clinical evaluation of advanced nerve guide conduits using a novel 3D in vitro testing model. *Int J Bioprinting* 4(1) 1-12.
- Stevenson G, Rehman S, Draper E, Hernández-Nava, E. Hunt J and Haycock JW (2016) Developing 3D human in vitro methods for evaluating novel porous titanium surfaces in orthopaedic applications. *Biotechnology & Bioengineering*. Vol.113(7), p.1586-1599.
- Hopper, AP, Dugan, JM, Gill, AA, Regan, EM, Haycock, JW, Kelly, S, May, PW, Claeysens, F (2016) Photochemically modified diamond-like carbon surfaces for neural interfaces. *Materials Science and Engineering C* 58(5725); 1199-1206.
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- Zilic L, Garner PE, Yu T, Roman S, Haycock JW, Wilshaw SP. (2015) An anatomical study of porcine peripheral and its potential in nerve tissue engineering. *Journal of Anatomy* 227(3); 302-314. Taylor CS,
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- Pateman C, Harding A, Glen A, Taylor C, Christmas C, Robinson P, Rimmer S, Boissonade F, Claeysens F, Haycock JW. (2015) Nerve guides manufactured from photocurable polymers to aid peripheral nerve repair. *Biomaterials* 49, 77–89.
- Kaewkhaw R, Scutt AM & Haycock JW (2012) A rapid method for the selective isolation of Schwann cells from adult nerve. *Nature Protocols* 7, 1996–2004.
- Kaewkhaw R, Scutt AM & Haycock JW (2011) Anatomical site influences the differentiation of adipose-derived stem cells for Schwann cell phenotype and function. *Glia* 59(5): 734-739.
- Daud MFB, Pawar KC, Claeysens F, Ryan AJ, Haycock JW (2012) An aligned 3D neuronal glial co-culture model for peripheral nerve studies. *Biomaterials* 33(25) 5901-5913.
- Murray-Dunning C & Haycock JW (2011). Three-dimensional alignment of Schwann cells using hydrolysable microfibre scaffolds: Strategies for peripheral nerve repair. *Methods Mol Biol* 695, 155-166.
- Haycock JW (2011). 3D Cell culture – a review of current techniques *Methods Mol Biol* 695, 1-16.
- Pateman C, Harding A, Glen A, Taylor C, Christmas C, Robinson P, Rimmer S, Boissonade F, Claeysens F, Haycock JW. (2015) Nerve guides manufactured from photocurable polymers to aid peripheral nerve repair. *Biomaterials* 49, 77–89.
- Hopper, A.P., Dugan, J.M, Gill, A.A., Fox, O.J.L., May, P.W., Haycock, J.W., Claeysens, F. (2014) Amine functionalized nanodiamond promotes cellular adhesion, proliferation and neurite outgrowth. *Biomedical Materials*, 9(4); 045009.
- Koroleva A, Gill AA, Ortega I, Haycock JW, Schlie S, Gittard SD, Chichkov BN, Claeysens, F (2012) Two-photon polymerization-generated and micromolding-replicated 3D scaffolds for peripheral neural tissue engineering applications. *Biofabrication* 3:1(2):025005

Acknowledgements

Dr Mehrie Behbehani
Mr Jonathan Field
Dr Dharaminder Singh
Dr Leyla Zilic
Dr Fauzi Daud
Dr Chris Pateman
Dr Adam Glen
Dr Caroline Taylor
Dr Adam Harding
Dr Ather Farook Khan
Dr Fred Claeysens
Prof Fiona Boissonade
Prof Ipsita Roy

